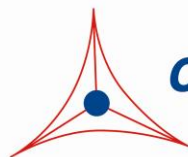

Product Manual

GFP ELISA Kit

Catalog Number

AKR-121	96 assays
AKR-121-5	5 x 96 assays

FOR RESEARCH USE ONLY
Not for use in diagnostic procedures



CELL BIOLABS, INC.
Creating Solutions for Life Science Research

Introduction

Green fluorescent protein (GFP) is a spontaneously fluorescent protein isolated from the pacific jellyfish, *Aequorea victoria*. It transduces the blue chemiluminescence into green fluorescent light. Since the molecular cloning of GFP cDNA and demonstration of GFP as a functional transgene, GFP has become a powerful tool with exciting applications in developmental, cell and molecular biology. GFP fluorescence is not species specific and can be expressed in bacteria, yeast, plant and mammalian cells. GFP can fuse with proteins of interest without interfering significantly with their assembly and function. Based on the structure of the GFP molecule, many GFP variants have been created with much improved fluorescence emission, or shifted excitation or emission spectra that are well suited for fluorescence microscopy and flow cytometry. Although GFP expression can be easily detected under a fluorescence microscope, GFP fluorescence intensity varies from cell to cell because of the heterogeneity nature of GFP expression. In order to quantitate the GFP expression in cells, FACS analysis is usually required, which is both expensive and time consuming.

Cell Biolabs' GFP ELISA Kit is an enzyme immunoassay developed for detection and quantitation of GFP or GFP fusion protein in cell or tissue samples. The quantity of GFP or its variants (including BFP, CFP and YFP) in an unknown sample is determined by comparing its absorbance with that of a known recombinant GFP standard curve. The kit has detection sensitivity limit of 30 pg/mL GFP. The kit also provides an efficient system for rapid quantitation of GFP lentivirus titer for both viral supernatant and purified virus. Each kit provides sufficient reagents to perform up to 96 assays including standard curve and GFP samples.

Related Products

1. AKR-120: GFP Quantitation Kit, Fluorometric
2. AKR-122: RFP ELISA Kit
3. STA-201: Recombinant EGFP

Kit Components

Box 1 (shipped at room temperature)

1. Anti-GFP Antibody Coated Plate (Part No. 212101): One 96-well strip plate (8 x 12).
2. Biotinylated Anti-GFP Antibody (1000X) (Part No. 212104): One 20 µL vial of biotinylated antibody recognizing jellyfish *Aequorea Victoria* GFP and its variants.
3. Streptavidin-Enzyme Conjugate (Part No. 310803): One 20 µL vial
4. Assay Diluent (Part No. 310804): One 50 mL bottle.
5. 10X Wash Buffer (Part No. 310806): One 100 mL bottle.
6. Substrate Solution (Part No. 310807): One 12 mL amber bottle.
7. Stop Solution (Part. No. 310808): One 12 mL bottle.

Box 2 (shipped on blue ice packs)

1. Recombinant GFP Standard (Part No. 212103): One 100 μL vial of 0.5 $\mu\text{g}/\text{mL}$ recombinant GFP in TBS plus BSA.

Materials Not Supplied

1. GFP Sample: cell or tissue lysate
2. 10 μL to 1000 μL adjustable single channel micropipettes with disposable tips
3. 50 μL to 300 μL adjustable multichannel micropipette with disposable tips
4. Multichannel micropipette reservoir
5. Microplate reader capable of reading at 450 nm (620 nm as optional reference wave length)

Storage

Upon receipt, aliquot and store recombinant GFP Standard at -80°C and avoid freeze/thaw. Store all other components at 4°C .

Preparation of Reagents

- 1X Wash Buffer: Dilute the 10X Wash Buffer Concentrate to 1X with deionized water. Stir to homogeneity.
- Anti-GFP Antibody and Streptavidin-Enzyme Conjugate: Immediately before use dilute the Biotinylated Anti-GFP antibody 1:1000 and the Streptavidin-Enzyme Conjugate 1:2000 with Assay Diluent. Do not store diluted solutions.

Preparation of GFP Standard

Prepare a dilution series of recombinant GFP standards in the concentration range of 0 ng/mL to 2 ng/mL in Assay Diluent (Table 1).

Standard Tubes	0.5 $\mu\text{g}/\text{mL}$ Recombinant GFP Standard (μL)	Assay Diluent (μL)	GFP ($\mu\text{g}/\text{mL}$)
1	4	996	2000
2	500 of Tube #1	500	1000
3	500 of Tube #2	500	500
4	500 of Tube #3	500	250
5	500 of Tube #4	500	125
6	500 of Tube #5	500	62.5
7	500 of Tube #6	500	31.2
8	0	500	0

Table 1. Preparation of samples for GFP Standard Curve

Assay Protocol

1. Prepare cell or tissue lysates containing GFP or GFP fusion protein.
Note: Because the ELISA kit has a linear range of 30 pg/mL to 2 ng/mL, we recommend using assay diluent to make series of 2-fold dilutions for each unknown sample.
2. Add 100 μ L of GFP sample or GFP standard to the Anti-GFP Antibody Coated Plate. Each GFP sample, GFP standard and blank should be assayed in duplicate.
3. Incubate at 37°C for at least 2 hours or 4°C overnight.
4. Wash microwell strips 3 times with 250 μ L 1X Wash Buffer per well with thorough aspiration between each wash. After the last wash, empty wells and tap microwell strips on absorbent pad or paper towel to remove excess 1X Wash Buffer.
5. Add 100 μ L of the diluted biotinylated anti-GFP antibody to each well.
6. Incubate at room temperature for 2 hours on an orbital shaker.
7. Wash the strip wells 3 times according to step 4 above.
8. Add 100 μ L of the diluted Streptavidin-Enzyme Conjugate to all wells.
9. Incubate at room temperature for 1 hour on an orbital shaker.
10. Wash the strip wells 3 times according to step 4 above. Proceed immediately to the next step.
11. Warm Substrate Solution to room temperature. Add 100 μ L of Substrate Solution to each well, including the blank wells. Incubate at room temperature on an orbital shaker. Actual incubation time may vary from 2-30 minutes.
Note: Watch plate carefully; if color changes rapidly, the reaction may need to be stopped sooner to prevent saturation.
12. Stop the enzyme reaction by adding 100 μ L of Stop Solution into each well, including the blank wells. Results should be read immediately (color will fade over time).
13. Read absorbance of each microwell on a spectrophotometer using 450 nm as the primary wave length.

Example of Results

The following figures demonstrate typical results with the GFP ELISA Kit. One should use the data below for reference only. This data should not be used to interpret actual results.

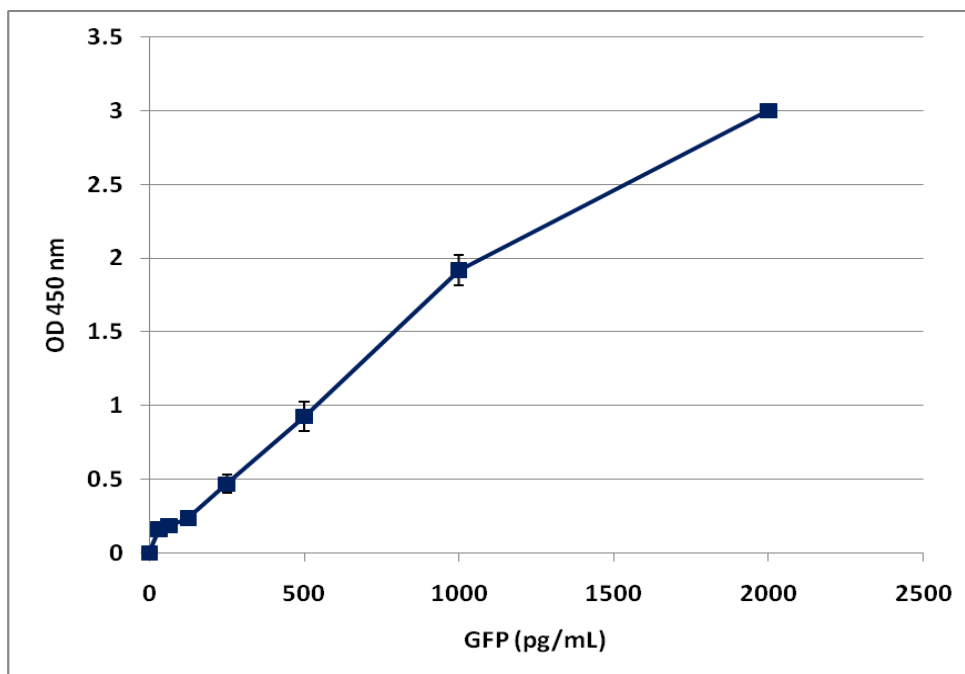


Figure 1: GFP ELISA Standard Curve.

References

1. Chalfie M. et al, *Science* **263**: 802-805, 1994.
2. Cormack B.P. et al, *Gene* **173**: 33-38, 1996.
3. Rizzuto R. et al, *Curr.Biol.* **6**:183-188 1996.

Recent Product Citations

1. Hata, Y. et al. (2022). A novel VCP modulator KUS121 exerts renoprotective effects in ischemia-reperfusion injury with retaining ATP and restoring ERAD-processing capacity. *Am J Physiol Renal Physiol.* **322**(5):F577-F586. doi: 10.1152/ajprenal.00392.2021.
2. Woods, V.M.A. et al. (2022). Targeting transgenic proteins to alpha granules for platelet-directed gene therapy. *Mol Ther Nucleic Acids.* doi: 10.1016/j.omtn.2021.12.038.
3. Czajka, M. et al. (2020). Mosaic Recombinant Adeno-associated Virus Vector rAAV/DJ/CAG for Targeted Gene Delivery to Melanoma Cells Metastasized to the Lung. *Anticancer Res.* **40**(8):4425-4444. doi: 10.21873/anticancer.14448.
4. Rodier, J.T. et al. (2019). Linear Polyethylenimine-DNA Nanoconstruct for Corneal Gene Delivery. *J Ocul Pharmacol Ther.* **35**(1):23-31. doi: 10.1089/jop.2018.0024.
5. Kwon, K.C. et al. (2019). An evaluation of microalgae as a recombinant protein oral delivery platform for fish using green fluorescent protein (GFP). *Fish Shellfish Immunol.* **87**:414-420. doi: 10.1016/j.fsi.2019.01.038.
6. Yang, B. et al. (2018). Comparative studies of the serum half-life extension of a protein via site-specific conjugation to a species-matched or -mismatched albumin. *Biomater Sci.* **6**(8):2092-2100. doi: 10.1039/c8bm00456k.
7. Obajemu, A.A. et al. (2017). IFN- λ 4 Attenuates Antiviral Responses by Enhancing Negative Regulation of IFN Signaling. *J Immunol.* **199**(11):3808-3820. doi: 10.4049/jimmunol.1700807.
8. Shrestha, R.P., and Hildebrand, M. et al (2017). Development of a silicon limitation inducible expression system for recombinant protein production in the centric diatoms *Thalassiosira*

- pseudonana and *Cyclotella cryptica*. *Microb Cell Fact.* **16**(1):145. doi: 10.1186/s12934-017-0760-3.
9. Fowler K.A., et al. (2017). Targeting the Canonical Nuclear Factor- κ B Pathway with a High-Potency IKK2 Inhibitor Improves Outcomes in a Mouse Model of Idiopathic Pneumonia Syndrome. *Biol Blood Marrow Transplant.* **23**(4):569-580. doi: 10.1016/j.bbmt.2017.01.083.
 10. Bruce, D.W. et al. (2017). Type 2 innate lymphoid cells treat and prevent acute gastrointestinal graft-versus-host disease. *J Clin Invest.* **127**(5):1813-1825. doi: 10.1172/JCI91816.
 11. Fowler, K.A. et al. (2017). Targeting the Canonical Nuclear Factor- κ B Pathway with a High-Potency IKK2 Inhibitor Improves Outcomes in a Mouse Model of Idiopathic Pneumonia Syndrome. *Biol Blood Marrow Transplant.* **23**(4):569-580. doi: 10.1016/j.bbmt.2017.01.083.
 12. Gordon, E.n D. et al. (2016). Alternative splicing of interleukin-33 and type 2 inflammation in asthma. *Proc Natl Acad Sci U S A.* **113**:8765-8770.
 13. Yang, B. et al. (2016). Site-specific albumination as an alternative to PEGylation for the enhanced serum half-life in vivo. *Biomacromolecules.* doi:10.1021/acs.biomac.6b00238.
 14. Vance, M. et al. (2016). AAV gene therapy for MPS1-associated corneal blindness. *Sci Rep.* doi:10.1038/srep22131.
 15. Johnson, K. A. et al. (2016). The Ebola Virus matrix protein, VP40, requires phosphatidylinositol 4, 5-bisphosphate (PI (4, 5) P2) for extensive oligomerization at the plasma membrane and viral egress. *Sci Rep.* **6**:19125.
 16. Chen, Z. et al. (2016). GADD45B mediates podocyte injury in zebrafish by activating the ROS-GADD45B-p38 pathway. *Cell Death Dis.* doi:10.1038/cddis.2015.300.
 17. Chen, C. C. et al. (2015). Changes in DNA methylation are associated with the development of drug resistance in cervical cancer cells. *Cancer Cell Int.* **15**:98.
 18. Borjan, B. et al. (2015). The Aplidin analogs PM01215 and PM02781 inhibit angiogenesis in vitro and in vivo. *BMC Cancer.* **15**:738.
 19. Gee, H. Y. et al. (2015). KANK deficiency leads to podocyte dysfunction and nephrotic syndrome. *J Clin Invest.* doi: 10.1172/JCI79504.
 20. Vemula, S. V. et al. (2015). HIV-1 induced Nuclear Factor IB (NF-IB) expression negatively regulates HIV-1 replication through interaction with the long terminal repeat region. *Viruses.* **7**:543-558.
 21. Zhang, Y. et al. (2015). Characterization of the promoter of grapevine vein clearing virus. *J Gen Virol.* **96**:165-169.
 22. Ott, L. & Carson, S. (2014). Immunological tools: Engaging students in the use and analysis of flow cytometry and enzyme-linked immunosorbent assay (ELISA). *Biochem Mol Biol Educ.* **42**:382-397.
 23. Fulton, L. M. et al. (2014). Altered T-cell entry and egress in the absence of Coronin 1A attenuates murine acute graft versus host disease. *Eur J Immunol.* **44**:1662-1671.
 24. Baez, A. et al. (2014). Production of recombinant protein by a novel oxygen-induced system in *Escherichia coli*. *Microb Cell Fact.* **13**:50.
 25. Huhtala, T. et al. (2014). Biodistribution and antitumor effect of Cetuximab-targeted lentivirus. *Nucl Med Biol.* **41**:77-83.
 26. Anyaegbu, C. C. et al. (2014). Chemotherapy enhances cross-presentation of nuclear tumor antigens. *PLoS One.* **9**:e107894.
 27. Sendra, L. et al. (2014). Low RNA translation activity limits the efficacy of hydrodynamic gene transfer to pig liver in vivo. *J Gene Med.* **16**:179-192.

28. Mango, R. et al. (2014). C-C Chemokine Receptor 5 on pulmonary mesenchymal cells promotes experimental metastasis via the induction of erythroid differentiation regulator 1. *Mol. Cancer. Res.* **12**:274-282.
29. Mitchell, A. et al. (2014). Promyelocytic leukemia protein is a cell-intrinsic factor inhibiting parvovirus DNA replication. *J. Virol.* **88**:925-936.
30. Coghill, J.M. et al. (2013). CC chemokine Receptor 8 potentiates donor treg survival and is critical for the prevention of murine graft-versus-host disease. *Blood.* **122**:825-836.

Warranty

These products are warranted to perform as described in their labeling and in Cell Biolabs literature when used in accordance with their instructions. THERE ARE NO WARRANTIES THAT EXTEND BEYOND THIS EXPRESSED WARRANTY AND CELL BIOLABS DISCLAIMS ANY IMPLIED WARRANTY OF MERCHANTABILITY OR WARRANTY OF FITNESS FOR PARTICULAR PURPOSE. CELL BIOLABS' sole obligation and purchaser's exclusive remedy for breach of this warranty shall be, at the option of CELL BIOLABS, to repair or replace the products. In no event shall CELL BIOLABS be liable for any proximate, incidental or consequential damages in connection with the products.

Contact Information

Cell Biolabs, Inc.
7758 Arjons Drive
San Diego, CA 92126
Worldwide: +1 858-271-6500
USA Toll-Free: 1-888-CBL-0505
E-mail: tech@cellbiolabs.com
www.cellbiolabs.com

©2013-2024: Cell Biolabs, Inc. - All rights reserved. No part of these works may be reproduced in any form without permissions in writing.