

## ZR-75-1/GFP Cell Line

**CATALOG NUMBER:** AKR-210

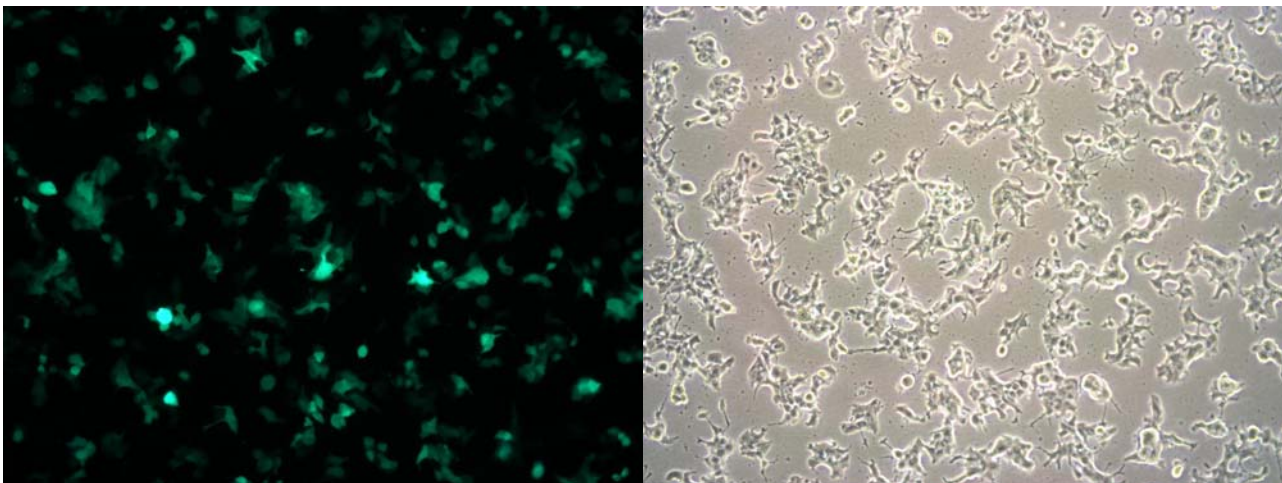
**STORAGE:** Liquid nitrogen

*Note: For best results begin culture of cells immediately upon receipt. If this is not possible, store at -80°C until first culture. Store subsequent cultured cells long term in liquid nitrogen.*

**QUANTITY & CONCENTRATION:** 1 mL,  $1 \times 10^6$  cells/mL in 70% DMEM, 20% FBS, 10% DMSO

### **Background**

The ZR-75-1 breast cancer cell line was derived in 1978 from a malignant ascetic effusion of pleural a 63 year old white female breast cancer patient. These cells exhibit epithelial morphology and form monolayers in culture. ZR-75-1 cells express the estrogen, oestrogen and progesterone receptors and are often used in hormone and tumourigenicity studies. Our ZR-75-1/GFP cell line stably expresses GFP; the gene was introduced using lentivirus.



**Figure 1.** ZR-75-1/GFP Cell Line. Left: GFP Fluorescence; Right: Phase Contrast.

### **Quality Control**

This cryovial contains at least  $1.0 \times 10^6$  ZR-75-1/GFP cells as determined by morphology, trypan-blue dye exclusion, and viable cell count. The ZR-75-1/GFP cells are tested free of microbial contamination.

### **Medium**

1. Culture Medium: RPMI-1640 or D-MEM (high glucose), 10% fetal bovine serum (FBS), 0.1 mM MEM Non-Essential Amino Acids (NEAA), 2 mM L-glutamine, 1% Pen-Strep.
2. Freeze Medium: 70% RPMI-1640 or DMEM, 20% FBS, 10% DMSO.

### **Methods**

### **Establishing ZR-75-1/GFP Cultures from Frozen Cells**

1. Place 10 mL of complete DMEM growth medium in a 50-mL conical tube. Thaw the frozen cryovial of cells within 1–2 minutes by gentle agitation in a 37°C water bath. Decontaminate the cryovial by wiping the surface of the vial with 70% (v/v) ethanol.
2. Transfer the thawed cell suspension to the conical tube containing 10 ml of growth medium.
3. Collect the cells by centrifugation at 1000 rpm for 5 minutes at room temperature. Remove the growth medium by aspiration.
4. Resuspend the cells in the conical tube in 15 mL of fresh growth medium by gently pipetting up and down.
5. Transfer the 15 mL of cell suspension to a T-75 tissue culture flask. Place the cells in a 37°C incubator at 5% CO<sub>2</sub>.
6. Monitor cell density daily. Cells should be passaged when the culture reaches 95% confluence.

*Note: ZR-75-1 growth doubling time is around 80 hrs.*

### **Warranty**

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***This product is for RESEARCH USE ONLY; not for use in diagnostic procedures.***

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