MDA-MB-436/RFP Cell Line

CATALOG NUMBER: AKR-252

STORAGE: **STORAGE:** Liquid nitrogen
Note: For best results begin culture of cells immediately upon receipt. If this is not possible, store at -80ºC until first culture. Store subsequent cultured cells long term in liquid nitrogen.

QUANTITY & CONCENTRATION: 1 mL, 1 x 10^6 cells/mL in 70% DMEM, 20% FBS, 10% DMSO

Background

The MDA-MB-436 breast cancer cell line was first derived from pleural fluid obtained from a 43-year-old breast cancer patient in 1976. MDA-MB-436 is pleomorphic and reacts intensely with anti tubulin antibody. *In vitro*, the MDA-MB-436 cell line has abundant activity in both the Boyden chamber chemoinvasion and chemotaxis assay. The MDA-MB-436 cell line is also able to grow on agarose, an indicator of transformation and tumorigenicity, and displays a relatively high colony forming efficiency. *In vivo*, the MDA-MB-436 cells form mammary fat pad tumors in nude mice. Our MDA-MB-436/RFP cell line stably expresses RFP and Puromycin resistant genes that were introduced using lentivirus.

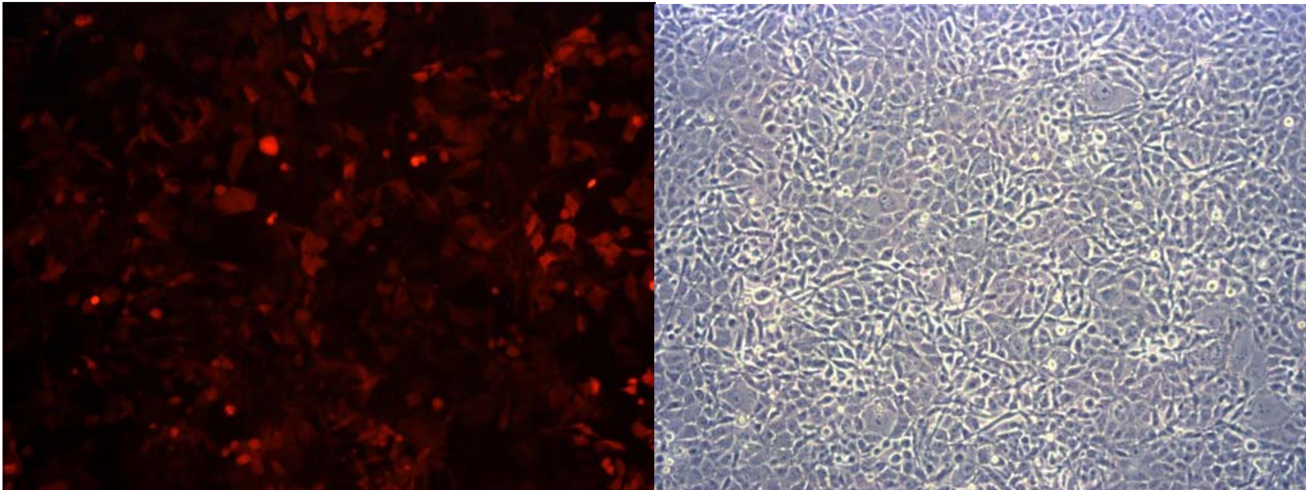

Figure 1. MDA-MB-436/RFP Cell Line. Left: RFP Fluorescence; Right: Phase Contrast.

Quality Control

This cryovial contains at least 1.0×10^6 MDA-MB-436/RFP cells as determined by morphology, trypan-blue dye exclusion, and viable cell count. The MDA-MB-436/RFP cells are tested free of microbial contamination.

Medium

1. Culture Medium: D-MEM (high glucose), 10% fetal bovine serum (FBS), 0.1 mM MEM Non-Essential Amino Acids (NEAA), 2 mM L-glutamine, 1% Pen-Strep.
2. Freeze Medium: 70% DMEM, 20% FBS, 10% DMSO.

Methods

Establishing MDA-MB-436/RFP Cultures from Frozen Cells

1. Place 10 mL of complete DMEM growth medium in a 50-mL conical tube. Thaw the frozen cryovial of cells within 1–2 minutes by gentle agitation in a 37°C water bath. Decontaminate the cryovial by wiping the surface of the vial with 70% (v/v) ethanol.
2. Transfer the thawed cell suspension to the conical tube containing 10 ml of growth medium.
3. Collect the cells by centrifugation at 1000 rpm for 5 minutes at room temperature. Remove the growth medium by aspiration.
4. Resuspend the cells in the conical tube in 15 mL of fresh growth medium by gently pipetting up and down.
5. Transfer the 15 mL of cell suspension to a T-75 tissue culture flask. Place the cells in a 37°C incubator at 5% CO_2.
6. Monitor cell density daily. Cells should be passaged when the culture reaches 95% confluence.

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