

---

Product Manual

# CytoSelect™ 24-Well Cell Migration Assay (8 µm, Colorimetric Format), Trial Size

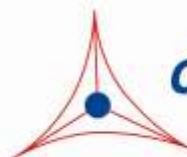
Catalog Number

CBA-100-T

4 assays

**FOR RESEARCH USE ONLY**  
Not for use in diagnostic procedures

---



**CELL BIOLABS, INC.**

*Creating Solutions for Life Science Research*

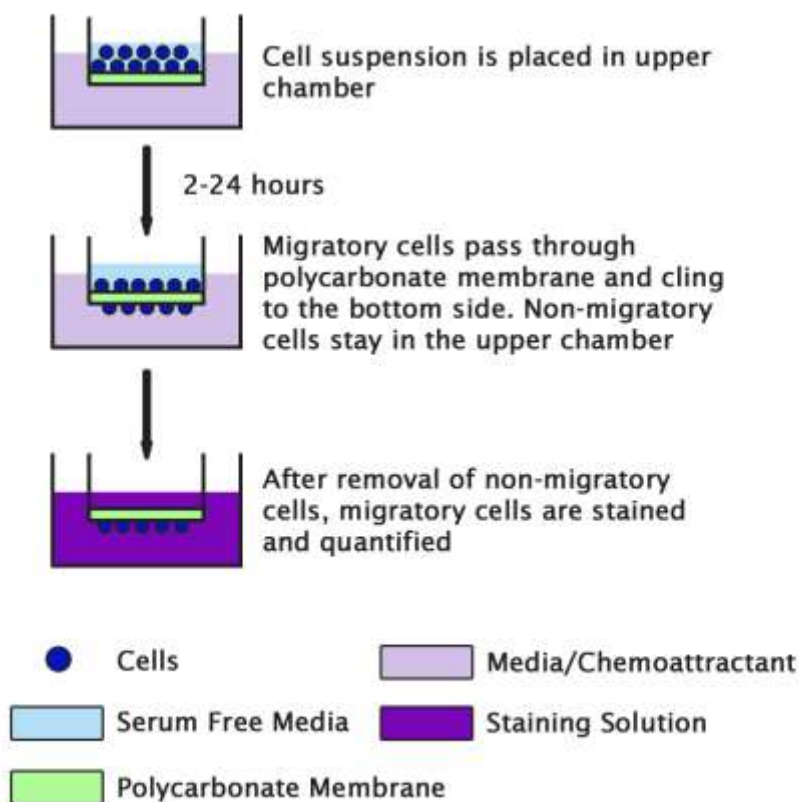
## **Introduction**

Cell migration is a highly integrated, multistep process that orchestrates embryonic morphogenesis, tissue repair and regeneration. It plays a pivotal role in the disease progression of cancer, mental retardation, atherosclerosis, and arthritis. The initial response of a cell to a migration-promoting agent is to polarize and extend protrusions in the direction of the attractant; these protrusions can consist of large, broad lamellipodia or spike-like filopodia. In either case, these protrusions are driven by actin polymerization and can be stabilized by extracellular matrix (ECM) adhesion or cell-cell interactions (via transmembrane receptors).

Cell Biolabs' CytoSelect™ Cell Migration Assay Kit utilizes polycarbonate membrane inserts (8 μm pore size) to assay the migratory properties of cells. This Trial Size kit contains sufficient reagents for the evaluation of 4 samples. The 8 μm pore size is optimal for epithelial and fibroblast cell migration. However, in the case of leukocyte chemotaxis, a smaller pore size (3 μm) is recommended.

## **Assay Principle**

The CytoSelect™ Cell Migration Assay Kit contains polycarbonate membrane inserts (8 μm pore size) in a 24-well plate. The membrane serves as a barrier to discriminate migratory cells from non-migratory cells. Migratory cells are able to extend protrusions towards chemoattractants (via actin cytoskeleton reorganization) and ultimately pass through the pores of the polycarbonate membrane. Finally, the cells are removed from the top of the membrane and the migratory cells are stained and quantified.



## **Related Products**

1. CBA-100-C: CytoSelect™ 24-Well Cell Migration and Invasion Assay (8µm, Colorimetric)
2. CBA-100-COL: CytoSelect™ 24-Well Cell Haptotaxis Assay (Collagen I, Colorimetric)
3. CBA-100-FN: CytoSelect™ 24-Well Cell Haptotaxis Assay (Fibronectin, Colorimetric)
4. CBA-101: CytoSelect™ 24-Well Cell Migration Assay (8µm, Fluorometric)
5. CBA-102: CytoSelect™ 24-Well Cell Migration Assay (5µm, Fluorometric)
6. CBA-103: CytoSelect™ 24-Well Cell Migration Assay (3µm, Fluorometric)
7. CBA-106: CytoSelect™ 96-Well Cell Migration Assay (8µm, Fluorometric)
8. CBA-110: CytoSelect™ 24-Well Cell Invasion Assay (Basement Membrane, Colorimetric)
9. CBA-125: Radius™ 24-Well Cell Migration Assay (Microscopy)
10. CBA-126: Radius™ 96-Well Cell Migration Assay (Microscopy)
11. CBA-130: CytoSelect™ 96-Well Cell Transformation Assay (Soft Agar Colony Formation)

## **Kit Components**

1. 24-well Migration Plate (Part No. 10001-T): One 24-well plate containing 4 cell culture inserts (8 µm pore size)
2. Cell Stain Solution (Part No. 11002-T): One 4 mL bottle
3. Extraction Solution (Part No. 11003-T): One 4 mL bottle
4. Cotton Swabs (Part No. 11004): 40 each
5. Forceps (Part No. 11005): One each

## **Materials Not Supplied**

1. Migratory cell lines
2. Cell culture medium
3. Serum free medium, such as DMEM containing 0.5% BSA, 2 mM CaCl<sub>2</sub> and 2 mM MgCl<sub>2</sub>
4. Cell culture incubator (37°C, 5% CO<sub>2</sub> atmosphere)
5. Light microscope
6. 96-well microtiter plate
7. Microtiter plate reader

## **Storage**

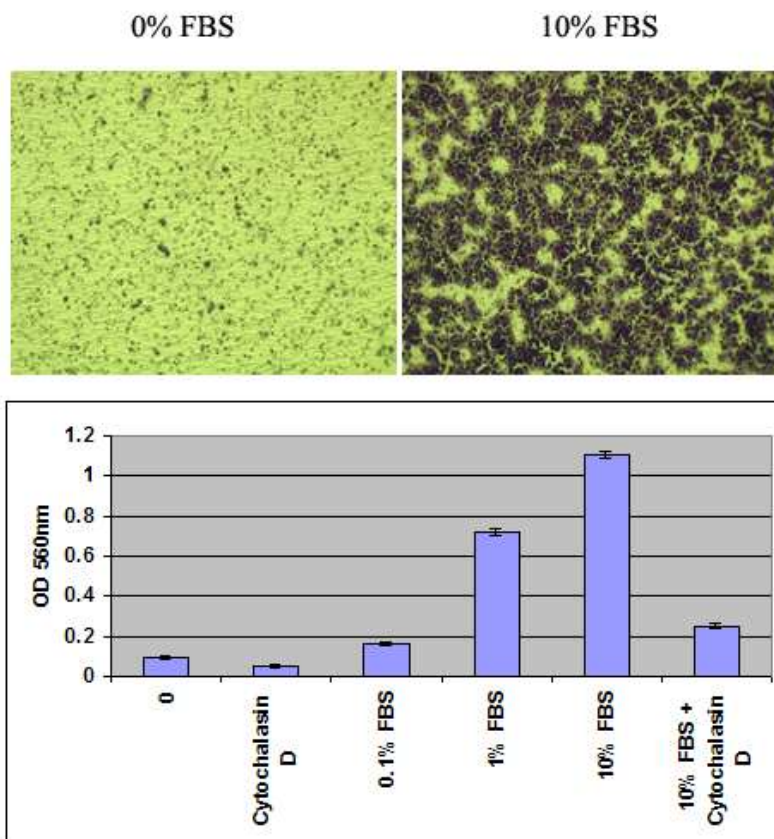
Store all components at 4°C.

## **Assay Protocol**

1. Under sterile conditions, allow the 24-well migration plate to warm up at room temperature for 10 minutes.
2. Prepare a cell suspension containing  $0.5-1.0 \times 10^6$  cells/ml in serum free media. Agents that inhibit or stimulate cell migration can be added directly to the cell suspension.  
*Note: Overnight starvation may be performed prior to running the assay*
3. Add 500  $\mu$ L of media containing 10% fetal bovine serum or desired chemoattractant(s) to the lower well of the migration plate.
4. Add 300  $\mu$ L of the cell suspension solution to the inside of each insert.
5. Incubate for 2-24 hours in a cell culture incubator.
6. Carefully aspirate the media from the inside of the insert. Wet the ends of 2-3 cotton-tipped swabs with water, flatten the ends of the swabs by pressing them against a clean hard surface, and gently swab the interior of the inserts to remove non-migratory cells. Take care not to puncture the polycarbonate membrane. Be sure to remove cells on the inside perimeter of the insert.
7. Transfer the insert to a clean well containing 400  $\mu$ L of Cell Stain Solution and incubate for 10 minutes at room temperature.
8. Gently wash the stained inserts several times in a beaker of water. Allow the inserts to air dry.
9. (optional) Count migratory cells with a light microscope under high magnification objective, with at least three individual fields per insert.
10. Transfer each insert to an empty well, adding 200  $\mu$ L of Extraction Solution per well, then incubating 10 minutes on an orbital shaker.
11. Transfer 100  $\mu$ L from each sample to a 96-well microtiter plate and measure the OD 560nm in a plate reader.

## Example of Results

The following figures demonstrate typical with the CytoSelect™ Cell Migration Assay Kit. One should use the data below for reference only. This data should not be used to interpret actual results.



**Figure 1. Human Fibrosarcoma HT-1080 Cell Migration.** HT-1080 cells were seeded at 150,000 cells/well and allowed to migrate toward FBS for 4 hrs in the presence or absence of 2  $\mu$ M Cytochalasin D. Migratory cells on the bottom of the polycarbonate membrane were stained (top panel picture) and quantified at OD 560nm after extraction (bottom panel figure).

## References

1. Ridley AJ, Schwartz MA, Burridge K, Firtel RA, Ginsberg MH, Borisy G, Parsons JT, Horwitz AR. (2003) *Science* **302**, 1704-9.
2. Horwitz R, Webb D. (2003) *Curr Biol.* **13**, R756-9.
3. Lauffenburger DA, Horwitz AF. (1996) *Cell* **84**, 359-369.

## **Recent Product Citations**

1. Liu, H. et al. (2021). Inhibition of USP11 sensitizes gastric cancer to chemotherapy via suppressing RhoA and Ras-mediated signaling pathways. *Clin Res Hepatol Gastroenterol*. doi: 10.1016/j.clinre.2021.101779.
2. Zacharias, N.M. et al. (2021). Prolyl Hydroxylase 3 Knockdown Accelerates VHL-Mutant Kidney Cancer Growth In Vivo. *Int J Mol Sci*. **22**(6):2849. doi: 10.3390/ijms22062849.
3. Ogawa, H. et al. (2021). Lenvatinib prevents liver fibrosis by inhibiting hepatic stellate cell activation and sinusoidal capillarization in experimental liver fibrosis. *J Cell Mol Med*. doi: 10.1111/jcmm.16363.
4. Zhu, S. et al. (2021). Ceramide kinase mediates intrinsic resistance and inferior response to chemotherapy in triple-negative breast cancer by upregulating Ras/ERK and PI3K/Akt pathways. *Cancer Cell Int*. **21**(1):42. doi: 10.1186/s12935-020-01735-5.
5. Royer-Pokora, B. et al. (2020). Comprehensive Biology and Genetics Compendium of Wilms Tumor Cell Lines with Different WT1 Mutations. *Cancers (Basel)*. **13**(1):E60. doi: 10.3390/cancers13010060.
6. Paik, E.S. et al. (2020). Preclinical assessment of the VEGFR inhibitor axitinib as a therapeutic agent for epithelial ovarian cancer. *Sci Rep*. **10**(1):4904. doi: 10.1038/s41598-020-61871-w.
7. Fouché, M. et al. (2020). Wound Healing Effects of Aloe muth-muth: In Vitro Investigations Using Immortalized Human Keratinocytes (HaCaT). *Biology (Basel)*. **9**(11):E350. doi: 10.3390/biology9110350.
8. Pinto, N. et al. (2020). Flavopiridol causes cell cycle inhibition and demonstrates anti-cancer activity in anaplastic thyroid cancer models. *PLoS One*. **15**(9):e0239315. doi: 10.1371/journal.pone.0239315.
9. Jun, J.H. et al. (2020). Regulation of Ras homolog family member G by microRNA-124 regulates proliferation and migration of human retinal pigment epithelial cells. *Sci Rep*. **10**(1):15420. doi: 10.1038/s41598-020-72360-5.
10. Chen, J. et al. (2020). Regulation of keratocyte phenotype and cell behavior by substrate stiffness. *ACS Biomater Sci Eng*. doi: 10.1021/acsbio.0c00510.
11. Kwon, S. et al. (2020). Biomarkers to quantify cell migration characteristics. *Cancer Cell Int*. **20**:217. doi: 10.1186/s12935-020-01312-w.
12. Oh, J.M. et al. (2020). U1 snRNP regulates cancer cell migration and invasion in vitro. *Nat Commun*. **11**(1):1. doi: 10.1038/s41467-019-13993-7.
13. Vay, S.U. et al. (2020). The impact of hyperpolarization-activated cyclic nucleotide-gated (HCN) and voltage-gated potassium KCNQ/Kv7 channels on primary microglia function. *J Neuroinflammation*. **17**(1):100. doi: 10.1186/s12974-020-01779-4.
14. Nyiramana, M.M. et al. (2020). Sea Hare Hydrolysate-Induced Reduction of Human Non-Small Cell Lung Cancer Cell Growth through Regulation of Macrophage Polarization and Non-Apoptotic Regulated Cell Death Pathways. *Cancers*. **12**:726. doi: 10.3390/cancers12030726.
15. Zheng, Q. et al. (2020). Cytotoxicity of amide-linked local anesthetics on melanoma cells via inhibition of Ras and RhoA signaling independent of sodium channel blockade. *BMC Anesthesiol*. **20**(1):43. doi: 10.1186/s12871-020-00957-4.
16. Chen, J. et al. (2020). Inhibition of arachidonate lipoxygenase12 targets lung cancer through inhibiting EMT and suppressing RhoA and NF- $\kappa$ B activity. *Biochem Biophys Res Commun*. pii: S0006-291X(20)30251-5. doi: 10.1016/j.bbrc.2020.01.166.

17. Chetty, S.S. et al. (2019). Human Umbilical Cord Wharton's Jelly-Derived Mesenchymal Stem Cells Labeled with Mn<sup>2+</sup> and Gd<sup>3+</sup> Co-Doped CuInS<sub>2</sub>-ZnS Nanocrystals for Multimodality Imaging in a Tumor Mice Model. *ACS Appl Mater Interfaces*. doi: 10.1021/acsami.9b19054.
18. Ali, R. et al. (2019). PARP1 blockade is synthetically lethal in XRCC1 deficient sporadic epithelial ovarian cancers. *Cancer Lett*. pii: S0304-3835(19)30538-5. doi: 10.1016/j.canlet.2019.10.035.
19. Rockfield, S. et al. (2019). Chronic iron exposure and c-Myc/H-ras-mediated transformation in fallopian tube cells alter the expression of EVI1, amplified at 3q26.2 in ovarian cancer. *Oncogenesis*. **8**(9):46. doi: 10.1038/s41389-019-0154-y.
20. Chu, Y. et al. (2019). Nudt21 regulates the alternative polyadenylation of Pak1 and is predictive in the prognosis of glioblastoma patients. *Oncogene*. doi: 10.1038/s41388-019-0714-9.
21. Wagner, G. et al. (2019). High Mobility Group Box 1 Protein in Osteoarthritic Knee Tissue and Chondrogenic Progenitor Cells: An Ex Vivo and In Vitro Study. *Cartilage*. 1947603519835897. doi: 10.1177/1947603519835897.
22. Chetty, S.S. et al. (2019). Noninvasive Tracking and Regenerative Capabilities of Transplanted Human Umbilical Cord-Derived Mesenchymal Stem Cells Labeled with I-III-IV Semiconducting Nanocrystals in Liver-Injured Living Mice. *ACS Appl Mater Interfaces*. **11**(9):8763-8778. doi: 10.1021/acsami.8b19953.
23. Poulard, C. et al. (2018). Increasing G9a automethylation sensitizes B acute lymphoblastic leukemia cells to glucocorticoid-induced death. *Cell Death Dis*. **9**(10):1038. doi: 10.1038/s41419-018-1110-z.
24. Wang, H.L. et al. (2018). Bulnesia sarmientoi Supercritical Fluid Extract Exhibits Necroptotic Effects and Anti-Metastatic Activity on Lung Cancer Cells. *Molecules*. **23**(12). pii: E3304. doi: 10.3390/molecules23123304.
25. Vay, S.U. et al. (2018). The plasticity of primary microglia and their multifaceted effects on endogenous neural stem cells in vitro and in vivo. *J Neuroinflammation*. **15**(1):226. doi: 10.1186/s12974-018-1261-y.
26. Choi, S.I. et al. (2017). Osteopontin production by TM4SF4 signaling drives a positive feedback autocrine loop with the STAT3 pathway to maintain cancer stem cell-like properties in lung cancer cells. *Oncotarget*. **8**(60):101284-101297. doi: 10.18632/oncotarget.21021.
27. Yamauchi, K., et al. (2017). Structure-Activity Relationships of Methylquercetin on Anti-migration and Anti-proliferation Activity in B16 Melanoma Cells. *Anticancer Res*. **37**(4):1575-1579.
28. Colden, M. et al. (2017). MicroRNA-466 inhibits tumor growth and bone metastasis in prostate cancer by direct regulation of osteogenic transcription factor RUNX2. *Cell Death Dis*. **8**(1):e2572. doi: 10.1038/cddis.2017.15.
29. Afghani, N. et al. (2017). Microtubule actin cross-linking factor 1, a novel target in glioblastoma. *Int. J. Oncol*. **50**:310-316.
30. Yu, X. M. et al. (2016). Notch1 signaling regulates the aggressiveness of differentiated thyroid cancer and inhibits SERPINE1 expression. *Clin Cancer Res*. doi: 10.1158/1078-0432.CCR-15-1749.

## **Warranty**

These products are warranted to perform as described in their labeling and in Cell Biolabs literature when used in accordance with their instructions. THERE ARE NO WARRANTIES THAT EXTEND BEYOND THIS EXPRESSED WARRANTY AND CELL BIOLABS DISCLAIMS ANY IMPLIED WARRANTY OF MERCHANTABILITY OR WARRANTY OF FITNESS FOR PARTICULAR PURPOSE. CELL BIOLABS' sole obligation and purchaser's exclusive remedy for breach of this warranty shall be, at the option of CELL BIOLABS, to repair or replace the products.

In no event shall CELL BIOLABS be liable for any proximate, incidental or consequential damages in connection with the products.

## **Contact Information**

Cell Biolabs, Inc.  
7758 Arjons Drive  
San Diego, CA 92126  
Worldwide: +1 858-271-6500  
USA Toll-Free: 1-888-CBL-0505  
E-mail: [tech@cellbiolabs.com](mailto:tech@cellbiolabs.com)  
[www.cellbiolabs.com](http://www.cellbiolabs.com)

©2013-2021: Cell Biolabs, Inc. - All rights reserved. No part of these works may be reproduced in any form without permissions in writing.