

---

Product Manual

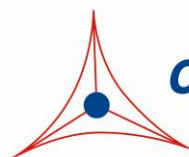
# CytoSelect™ 96-Well Cell Migration Assay (8 µm, Fluorometric Format)

## Catalog Number

CBA-106	96 assays
CBA-106-5	5 x 96 assays

**FOR RESEARCH USE ONLY**  
**Not for use in diagnostic procedures**

---



**CELL BIOLABS, INC.**  
*Creating Solutions for Life Science Research*

## **Introduction**

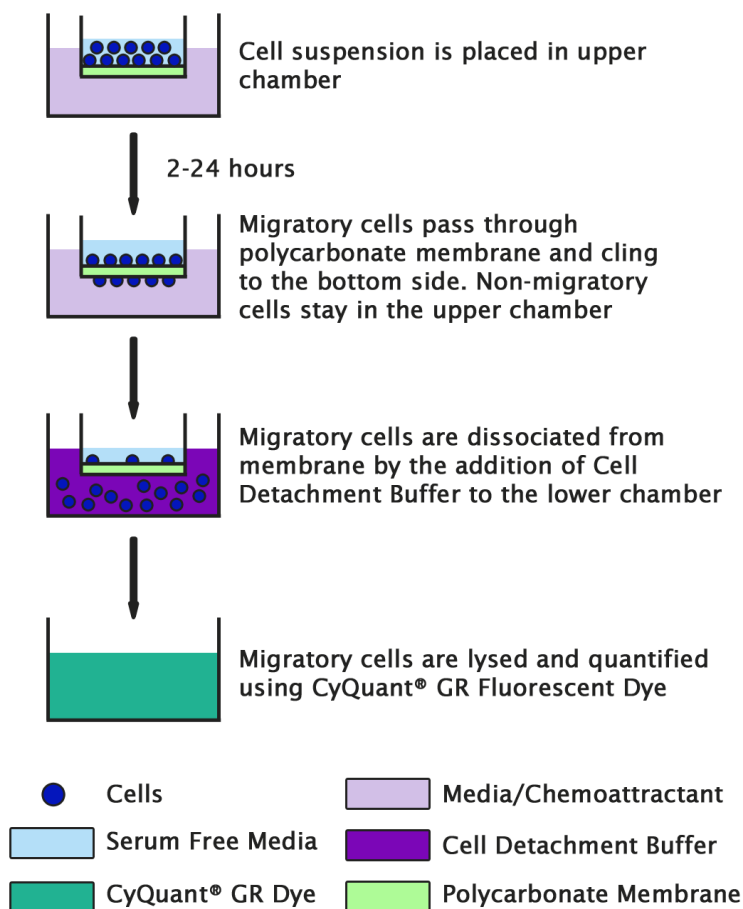
Cell migration is a highly integrated, multistep process that orchestrates embryonic morphogenesis, tissue repair and regeneration. It plays a pivotal role in the disease progression of cancer, atherosclerosis, and arthritis. The initial response of a cell to a migration-promoting agent is to polarize and extend protrusions in the direction of the attractant; these protrusions can consist of large, broad lamellipodia or spike-like filopodia. In either case, these protrusions are driven by actin polymerization and can be stabilized by extracellular matrix (ECM) adhesion or cell-cell interactions.

Cell Biolabs CytoSelect™ 96-well Cell Migration Assay Kit utilizes a polycarbonate membrane plate (8 µm pore size) to assay the migratory properties of cells. The kit does not require you to prelabel the cells with Calcein AM or remove non-migratory cells (i.e., cotton swabbing). Any migratory cells are first dissociated from the membrane, then lysed and detected with CyQuant® GR Dye.

Cell Biolabs CytoSelect™ 96-well Cell Migration Assay Kit provides a robust system for the quantitative determination of cell migration. The kit contains sufficient reagents for the evaluation of 96 samples. The 8 µm pore size is optimal for epithelial and fibroblast cell migration. However, in the case of leukocyte chemotaxis, a smaller pore size (3 µm) is recommended.

The CytoSelect™ Cell Migration Assay Kit contains a polycarbonate membrane chamber (8 µm pore size) in a 96-well plate. The membrane serves as a barrier to discriminate migratory cells from non-migratory cells. Migratory cells are able to extend protrusions towards chemoattractants (via actin cytoskeleton reorganization) and ultimately pass through the pores of the polycarbonate membrane. These migratory cells are then dissociated from the membrane and subsequently detected with CyQuant® GR Dye.

## **Assay Principle**



## **Related Products**

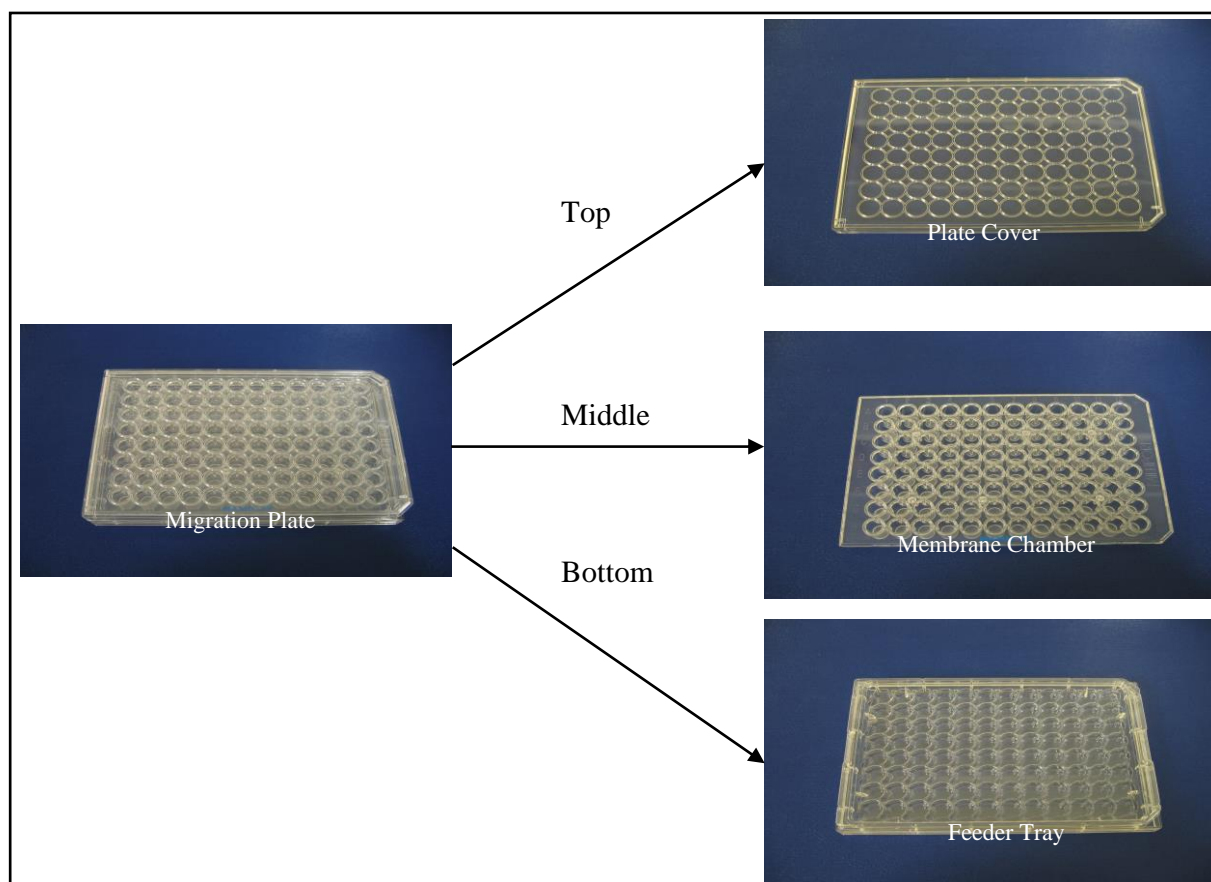
1. CBA-101: CytoSelect™ 24-Well Cell Migration Assay (8µm, Fluorometric)
2. CBA-102: CytoSelect™ 24-Well Cell Migration Assay (5µm, Fluorometric)
3. CBA-103: CytoSelect™ 24-Well Cell Migration Assay (3µm, Fluorometric)
4. CBA-104: CytoSelect™ 96-Well Cell Migration Assay (3µm, Fluorometric)
5. CBA-105: CytoSelect™ 96-Well Cell Migration Assay (5µm, Fluorometric)

## **Kit Components (shipped at room temperature)**

1. 96-well Cell Migration Plate (Part No. 10601): One sterile 96-well plate (see Figure 1 for components)
2. 96-well Cell Harvesting Tray (Part No. 10402): One 96-well tray
3. Cell Detachment Solution (Part No. 10403): One 20 mL bottle
4. 4X Lysis Buffer (Part No. 10404): One 10 mL bottle
5. CyQuant® GR Dye (Part No. 10105): One 75 µL tube

## **Materials Not Supplied**

1. Migratory cell lines
2. Cell culture medium
3. Serum free medium, such as DMEM containing 0.5% BSA, 2 mM  $\text{CaCl}_2$  and 2 mM  $\text{MgCl}_2$
4. FBS or desired chemoattractant
5. Cell culture incubator (37°C, 5%  $\text{CO}_2$  atmosphere)
6. Light microscope
7. 96-well plate suitable for a fluorescence plate reader
8. Fluorescence plate reader



**Figure 1: Components of the 96-well Cell Migration Plate.**

## **Storage**

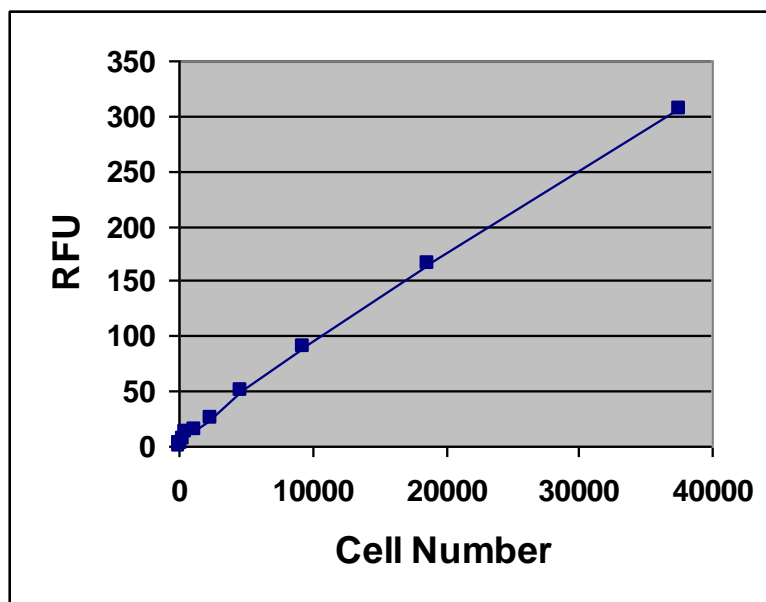
Store all components at 4°C.

## **Assay Protocol**

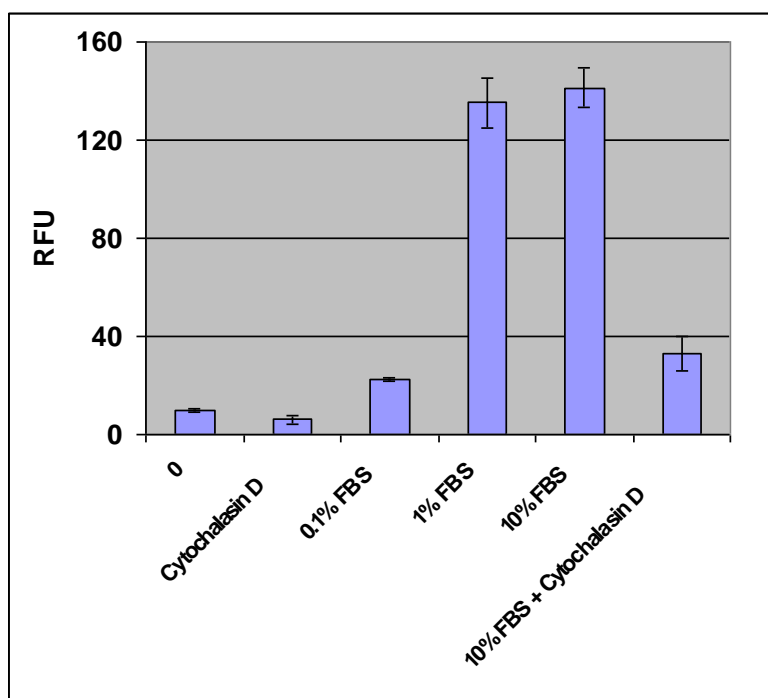
1. Allow the 96-well Migration Plate to warm up at room temperature for 10 minutes.
2. Prepare a cell suspension containing  $0.1\text{--}1.0 \times 10^6$  cells/ml in serum free media. Agents that inhibit or stimulate cell migration can be added directly to the cell suspension.  
(Note: Overnight starvation may be performed prior to running the assay)
3. Under sterile conditions, separate the cover and membrane chamber from the 96-well Migration Plate.
4. Add 150  $\mu\text{L}$  of media containing 10% fetal bovine serum or desired chemoattractant(s) to the wells of the feeder tray.
5. Place the membrane chamber back into the feeder tray (containing chemoattractant solution).  
**Ensure no bubbles are trapped under the membrane.**
6. Gently mix the cell suspension (without chemoattractant) from step 2 and add 100  $\mu\text{L}$  to the membrane chamber.
7. Finally, cover the plate and transfer to a cell culture incubator for 2-24 hours.
8. Just prior to the end of the incubation, pipette 150  $\mu\text{L}$  of prewarmed Cell Detachment Solution into wells of the clean, 96-Well Cell Harvesting Tray (provided).
9. Carefully remove the 96-well Migration Plate from the incubator. Separate the membrane chamber from the feeder tray.
10. Remove the cells/media from the top side of the membrane chamber by aspirating or inverting. Place the membrane chamber into the Cell Harvesting Tray containing 150  $\mu\text{L}$  of Cell Detachment Solution (step 8). Incubate 30 minutes at 37°C.
11. Completely dislodge the cells from the underside of the membrane by gently tilting the membrane chamber several times in the Cell Detachment Solution.
12. Prepare sufficient 4X Lysis Buffer/CyQuant® GR dye solution for all samples by diluting the dye 1:75 in 4X Lysis Buffer (for example, add 5  $\mu\text{L}$  dye to 370  $\mu\text{L}$  of 4X Lysis Buffer).
13. Add 50  $\mu\text{L}$  of 4X Lysis Buffer/CyQuant® GR dye solution to each well (already containing 150  $\mu\text{L}$  of Cell Detachment Solution). Incubate 20 minutes at room temperature.
14. Transfer 150  $\mu\text{L}$  of the mixture to a 96-well plate suitable for fluorescence measurement. Read the fluorescence with a fluorescence plate reader at 480 nm/520 nm.

## **Example of Results**

The following figures demonstrate typical with the CytoSelect™ Cell Migration Assay Kit. One should use the data below for reference only. This data should not be used to interpret actual results.



**Figure 2: Quantitation of Human HT-1080.** HT-1080 cells were titrated in Cell Detachment Buffer, then subsequently lysed and detected with 4X Lysis Buffer/Cyquant® GR Dye (150  $\mu$ L cell suspension was mixed with 50  $\mu$ L of 4X Lysis Buffer/dye).



**Figure 3: HT-1080 Chemotaxis.** HT-1080 cells were allowed to migrate toward FBS for 4 hrs in the presence or absence of 2  $\mu$ M Cytochalasin D, 30,000 cells were used in each assay. Migratory cells on the bottom of the polycarbonate membrane were detached and quantified by CyQuant® GR Dye as described in the Assay Protocol.

## **References**

1. Ridley AJ, Schwartz MA, Burridge K, Firtel RA, Ginsberg MH, Borisy G, Parsons JT, Horwitz AR. (2003) *Science* **302**, 1704-9.
2. Horwitz R, Webb D. (2003) *Curr Biol.* **13**, R756-9.
3. Lauffenburger DA, Horwitz AF. (1996) *Cell* **84**, 359-369.

## **Recent Product Citations**

1. Popławski, P. et al. (2023). Renal cancer secretome induces migration of mesenchymal stromal cells. *Stem Cell Res Ther.* **14**(1):200. doi: 10.1186/s13287-023-03430-4.
2. Suwatthanarak, T. et al. (2023). Screening of EWI-2-Derived Peptides for Targeting Tetraspanin CD81 and Their Effect on Cancer Cell Migration. *Biomolecules.* **13**(3):510. doi: 10.3390/biom13030510.
3. López-Moncada, F. et al. (2022). SPARC Induces E-Cadherin Repression and Enhances Cell Migration through Integrin  $\alpha\beta3$  and the Transcription Factor ZEB1 in Prostate Cancer Cells. *Int J Mol Sci.* **23**(11):5874. doi: 10.3390/ijms23115874.
4. Avolio, E. et al. (2021). Secreted Protein Acidic and Cysteine Rich Matricellular Protein is Enriched in the Bioactive Fraction of the Human Vascular Pericyte Secretome. *Antioxid Redox Signal.* **34**(15):1151-1164. doi: 10.1089/ars.2019.7969.
5. Chen, J. et al. (2021). NOX5 mediates the crosstalk between tumor cells and cancer-associated fibroblasts via regulating cytokine network. *Clin Transl Med.* **11**(8):e472. doi: 10.1002/ctm2.472.
6. Lee, S. et al. (2021). Discovery of novel potent migrastatic Thiazolo[5,4-b]pyridines targeting Lysyl-tRNA synthetase (KRS) for treatment of Cancer metastasis. *Eur J Med Chem.* doi: 10.1016/j.ejmech.2021.113405.
7. Yi, S.W. et al. (2021). Dilation-Responsive Microshape Programing Prevents Vascular Graft Stenosis. *Small.* doi: 10.1002/sml.202007297.
8. Avolio, E. et al. (2020). Secreted Protein Acidic and Cysteine Rich Matricellular Protein Is Enriched in the Bioactive Fraction of the Human Vascular Pericyte Secretome. *Antioxid Redox Signal.* doi: 10.1089/ars.2019.7969.
9. Ma, L. et al. (2020). Targeted MEK inhibition by cobimetinib enhances doxorubicin's efficacy in osteosarcoma models. *Biochem Biophys Res Commun.* **529**(3):622-628. doi: 10.1016/j.bbrc.2020.06.082.
10. Vílchez, J.I. et al. (2020). DNA demethylases are required for myo-inositol-mediated mutualism between plants and beneficial rhizobacteria. *Nat Plants.* doi: 10.1038/s41477-020-0707-2.
11. Ntogwa, M. et al. (2020). Schwann cell-derived CXCL1 contributes to human immunodeficiency virus type 1 gp120-induced neuropathic pain by modulating macrophage infiltration in mice. *Brain Behav Immun.* doi: 10.1016/j.bbi.2020.03.027.
12. Umezu, K. et al. (2020). Stromal cell-derived factor 1 regulates in vitro sperm migration towards the cumulus-oocyte complex in cattle. *PLoS One.* **15**(4):e0232536. doi: 10.1371/journal.pone.0232536.
13. Morcillo, R.J. et al. (2019). Rhizobacterium-derived diacetyl modulates plant immunity in a phosphate-dependent manner. *EMBO J.* doi: 10.15252/embj.2019102602.
14. Sakurai, K. et al. (2019). CD36 expression on oral squamous cell carcinoma cells correlates with enhanced proliferation and migratory activity. *Oral Dis.* doi: 10.1111/odi.13210.
15. Anitua, E. et al. (2019). A novel protein-based autologous topical serum for skin regeneration. *J Cosmet Dermatol.* doi: 10.1111/jocd.13075.



16. Muraguchi, T. et al. (2019). IGF-1R deficiency in human keratinocytes disrupts epidermal homeostasis and stem cell maintenance. *Journal of Dermatological Science*. doi: 10.1016/j.jdermsci.2019.05.001.
17. Tian, S. et al. (2018). The prognostic roles of circulating ALDH1+ tumor cell in the patients with non-small cell lung cancer. *Biosci Rep*. **38**(5). pii: BSR20180914. doi: 10.1042/BSR20180914.
18. Li, Z. et al. (2018). The transcriptional coactivator WBP2 primes triple-negative breast cancer cells for responses to Wnt signaling via the JNK/Jun kinase pathway. *J Biol Chem*. **293**(52):20014-20028. doi: 10.1074/jbc.RA118.005796.
19. Fedyakova, E. et al. (2018). An autologous protein gel for soft tissue augmentation: in vitro characterization and clinical evaluation. *J Cosmet Dermatol*. doi: 10.1111/jocd.12771.
20. Zhang, M. et al. (2018). AIBP reduces atherosclerosis by promoting reverse cholesterol transport and ameliorating inflammation in apoE<sup>-/-</sup> mice. *Atherosclerosis*. **273**:122-130. doi: 10.1016/j.atherosclerosis.2018.03.010.
21. Chen, H. et al. (2017). The Exonization and Functionalization of an Alu-J Element in the Protein Coding Region of Glycoprotein Hormone Alpha Gene Represent a Novel Mechanism to the Evolution of Hemochorial Placentation in Primates. *Mol Biol Evol*. **34**(12):3216-3231. doi: 10.1093/molbev/msx252.
22. Barazeghi, E. et al. (2017). A role for TET2 in parathyroid carcinoma. *Endocr. Relat. Cancer* **24**(7):329-336.
23. Jung, H-S. et al (2017). Monoclonal antibodies against autocrine motility factor suppress gastric cancer. *Oncology Letters*. **13** (6): 4925-4932.
24. Ben-David, U. et al. (2016). The landscape of chromosomal aberrations in breast cancer mouse models reveals driver-specific routes to tumorigenesis. *Nat Commun*. doi:10.1038/ncomms12160.
25. Adam, M. G. et al. (2015). SIAH ubiquitin ligases regulate breast cancer cell migration and invasion independent of the oxygen status. *Cell Cycle*. **14**:3734-3747.
26. Ranchoux, B. et al. (2015). Endothelial-to-Mesenchymal Transition in Pulmonary Hypertension. *Circulation*. **131**:1006-1018.
27. Li, X. et al. (2013). Activation of thromboxane A2 receptor (TP) increases the expression of monocyte chemoattractant protein -1 (MCP-1)/chemokine (C-C motif) ligand 2 (CCL2) and recruits macrophages to promote invasion of lung cancer cells. *PLoS One*. **8**(1):e54073. doi: 10.1371/journal.pone.0054073.
28. Rosenblum, S. et al. (2012). Timing of intra-arterial neural stem cell transplantation after hypoxia-ischemia influences cell engraftment, survival, and differentiation. *Stroke*. **43**:1624-1631.
29. Aftab, B.T. et al. (2011). Itraconazole inhibits angiogenesis and tumor growth in non-small cell lung cancer. *Cancer Res*. **71**:6764-6772.
30. Andres, R.H. et al. (2011). The CCR2/CCL2 interaction mediates the transendothelial recruitment of intravascularly delivered neural stem cells to the ischemic brain. *Stroke* **42**:2923-2931.

## **License Information**

This product is provided under an intellectual property license from Life Technologies Corporation. The purchase of this product conveys to the buyer the non-transferable right to use the purchased product and components of the product only in research conducted by the buyer (whether the buyer is an academic or for-profit entity). The sale of this product is expressly conditioned on the buyer not using the product or its components, or any materials made using the product or its components, in any activity to generate revenue, which may include, but is not limited to use of the product or its components: (i) in manufacturing; (ii) to provide a service, information, or data in return for payment;



(iii) for therapeutic, diagnostic or prophylactic purposes; or (iv) for resale, regardless of whether they are resold for use in research. For information on purchasing a license to this product for purposes other than as described above, contact Life Technologies Corporation, 5791 Van Allen Way, Carlsbad CA 92008 USA or [outlicensing@lifetech.com](mailto:outlicensing@lifetech.com).

## **Warranty**

These products are warranted to perform as described in their labeling and in Cell Biolabs literature when used in accordance with their instructions. THERE ARE NO WARRANTIES THAT EXTEND BEYOND THIS EXPRESSED WARRANTY AND CELL BIOLABS DISCLAIMS ANY IMPLIED WARRANTY OF MERCHANTABILITY OR WARRANTY OF FITNESS FOR PARTICULAR PURPOSE. CELL BIOLABS' sole obligation and purchaser's exclusive remedy for breach of this warranty shall be, at the option of CELL BIOLABS, to repair or replace the products. In no event shall CELL BIOLABS be liable for any proximate, incidental or consequential damages in connection with the products.

## **Contact Information**

Cell Biolabs, Inc.  
5628 Copley Drive  
San Diego, CA 92111  
Worldwide: +1 858-271-6500  
USA Toll-Free: 1-888-CBL-0505  
E-mail: [tech@cellbiolabs.com](mailto:tech@cellbiolabs.com)  
[www.cellbiolabs.com](http://www.cellbiolabs.com)

©2004-2024: Cell Biolabs, Inc. - All rights reserved. No part of these works may be reproduced in any form without permissions in writing.