

---

Product Manual

# Cell Contraction Assay

Catalog Number

CBA-201

24 assays

**FOR RESEARCH USE ONLY**  
Not for use in diagnostic procedures

---



**CELL BIOLABS, INC.**  
*Creating Solutions for Life Science Research*

## **Introduction**

Wound healing comprises of three processes: epithelialization, connective tissue deposition, and contraction. The contraction process is believed to be mediated by specialized fibroblasts called myofibroblasts. Three-dimensional collagen gels have been widely used in fibroblast contraction studies.

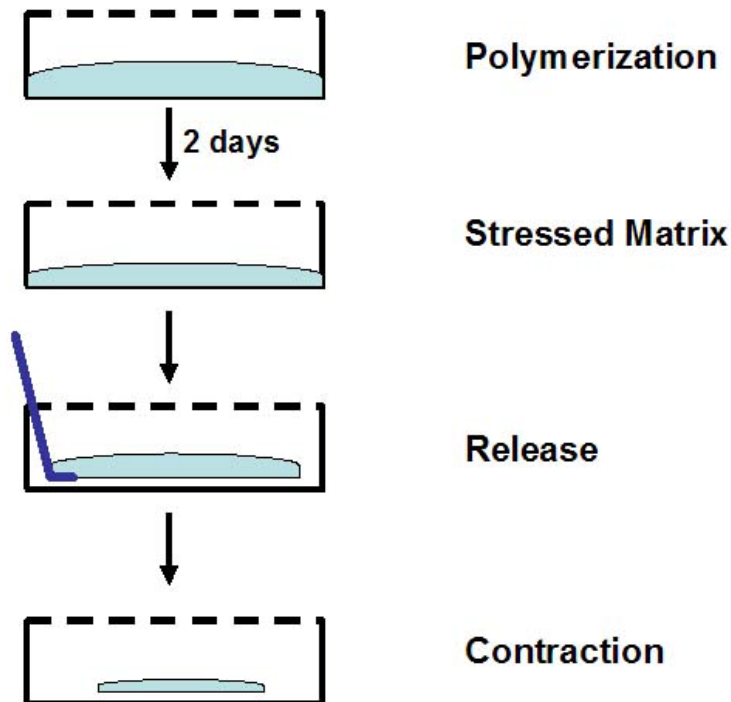
There are several different culture models to study the ability of fibroblasts to reorganize and contract collagen matrices in vitro. In the floating contraction model, a freshly polymerized collagen matrix containing cells is released from the culture dish and allowed to float in culture medium, and contraction occurs in the absence of external mechanical load and without appearance of stress fibers in the cells. In the attached model, a polymerized collagen matrix containing cells remains attached to the culture dish during contraction. Mechanical tension develops during contraction, and cellular stress fibers assemble. The two-step model combines an initial period of attached matrix contraction leading to mechanical loading, followed by release of the matrices, resulting in mechanical unloading and further contraction as mechanical stress dissipates.

The signaling mechanisms used by fibroblasts to regulate collagen matrix contraction depend on whether the cells are mechanically loaded or unloaded at the time that contraction is initiated as well as on the growth factor used to initiate contraction. For instance, stimulation of fibroblasts by lysophosphatidic acid (LPA) but not by platelet-derived growth factor (PDGF) causes robust force generation in restrained matrices, whereas LPA and PDGF stimulate floating matrix contraction equally well.

3D collagen matrix has also been used in the studies of integrin signaling, cell apoptosis and cytoskeleton reorganization. Since three-dimensional matrix adhesions differ in structure, localization, and function from two-dimensional adhesions; and therefore, three-dimensional cell-matrix interactions may be more relevant biologically.

Cell Biolabs' Collagen-based Contraction Assay Kit provides a simple system to assess cell contractivity in vitro and screen cell contraction mediators. Each kit provides sufficient quantities to perform up to 24 assays in a 24-well plate. The kit can also be used for culturing cells in a 3D collagen matrix.

## Assay Principle



## Kit Components

1. Collagen Solution (Part No. 20101): One 10 mL bottle of sterile bovine Type I Collagen at 3.0 mg/mL
2. Neutralization Solution (Part No. 20102): One 0.5 mL tube
3. 5X DMEM Medium (Part No. 20103): One 5 mL bottle
4. 5X PBS (Part No. 20104): One 5 mL bottle
5. 100X Cell Contraction Inhibitor (Part No. 20105): One 1 mL tube of 1M 2, 3-Butanedione Monoxime (BDM) in DMSO

## Materials Not Supplied

1. Cells such as fibroblasts
2. Cell culture medium
3. 37°C Incubator, 5% CO<sub>2</sub> atmosphere
4. Sterile Spatula
5. Light microscope
6. Ruler

## Storage

Store all components at 4°C.

## **Preparation of Collagen Gel Working Solution**

This kit is designed for samples in a 24-well plate, and may be modified accordingly to suit other culture plate sizes. Keep all solutions ON ICE the entire time.

***Important Note: Be sure to pipet all volumes carefully with well-calibrated pipettes. Volumes of each reagent are critical for collagen polymerization.***

1. In a cold sterile tube, add the desired amount of Collagen Solution according to the table below. Next, add 5X DMEM medium or 5X PBS to the tube and mix well.
2. Add Neutralization solution, IMMEDIATELY mix and keep the Collagen Gel Working Solution on ice.

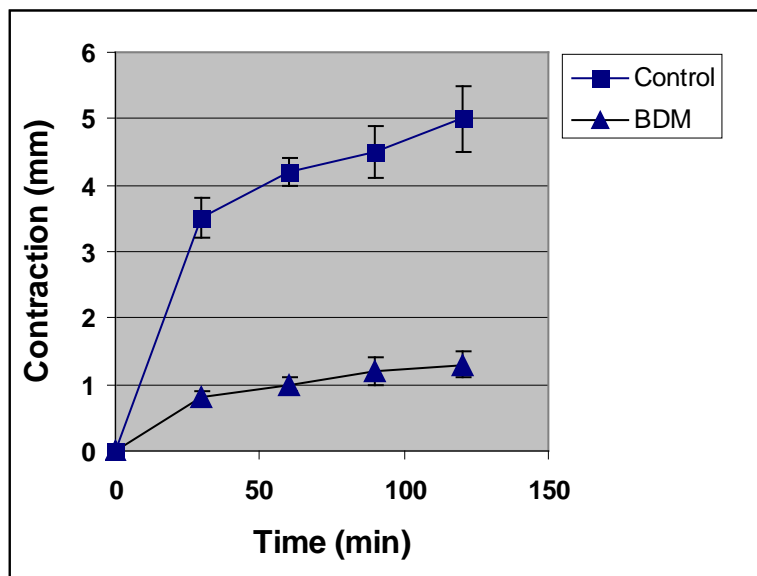
<b>Reagents</b>	<b>6 wells</b>	<b>12 wells</b>	<b>24 wells</b>
Collagen Solution	2.385 mL	4.77 mL	9.54 mL
5X Medium or PBS	615 $\mu$ L	1.23 mL	2.46 mL
Neutralization Solution	85 $\mu$ L	170 $\mu$ L	340 $\mu$ L
<b>Total</b>	<b>3.085 mL</b>	<b>6.17 mL</b>	<b>12.34 mL</b>

## **Assay Protocol (Two-Step Collagen Contraction Model)**

1. Harvest cells and resuspend in desired medium at  $2-5 \times 10^6$  cells/mL.
2. Prepare the collagen lattice by mixing 2 parts of cell suspension and 8 parts of cold Collagen Gel Working Solution.
3. Add 0.5 mL of the cell-collagen mixture per well in a 24-well plate, incubate 1 hr at 37°C.
4. After collagen polymerization, 1.0 mL of culture medium is added atop each collagen gel lattice.
5. Cultures are incubated for two days, during which stress develops. Before releasing the stressed matrix, cells may be treated with contraction mediators, such as 10 mM BDM. To initiate contraction, gently release collagen gels from the sides of the culture dishes with a sterile spatula.
6. The collagen gel size change (contraction index) can be measured at various times with a ruler or quantified with image analysis software, such as NIH Image or Image Pro Plus.

## Example of Results

The following figure demonstrates typical contraction results using the Cell Contraction Assay. One should use the data below for reference only. This data should not be used to interpret actual results.



**Figure 1. Contraction inhibition by BDM.**  $0.5 \times 10^6$  COS-7 cells in 0.5 mL collagen gel lattice were cultured for two days. Before initiation of contraction, cells were pretreated with 10 mM BDM for 1 hr. The change of gel size (diameter) in millimeters was measured with a ruler at various times after release.

## References

1. Martin, P. (1997) *Science* **276**, 75-81
2. Bell, E., Ivarsson, B., and Merrill, C. (1979) *Proc. Natl. Acad. Sci. U. S. A.* **76**, 1274-1278
3. Stupak, D., and Harris, A. K. (1982) *Dev. Biol.* **90**, 383-398
4. Mochitate, K., Pawelek, P., and Grinnell, F. (1991) *Exp. Cell Res.* **193**, 198-207
5. Tian, B., Lessan, K., Kahm, J., Kleidon, J., and Henke, C. (2002) *J. Biol. Chem.* **277**, 24667-24675

## Recent Product Citations

1. Bravo, D.D. et al. (2018). An impedance-based cell contraction assay using human primary smooth muscle cells and fibroblasts. *J Pharmacol Toxicol Methods.* **89**:47-53. doi: 10.1016/j.vascn.2017.10.006.
2. Schafer, S. et al. (2017). IL-11 is a crucial determinant of cardiovascular fibrosis. *Nature.* **552**(7683):110-115. doi: 10.1038/nature24676.
3. Liu, L. et al. (2017). Two potent alpha3/5 conotoxins from piscivorous *Conus achatinus*. *Acta Biochim Biophys Sin (Shanghai).* **39**(6):438-44.
4. Jung, J.E. et al. (2017). Local myogenic pulp-derived cell injection enhances craniofacial muscle regeneration in vivo. *Orthod Craniofac Res.* **20**(1):35-43. doi: 10.1111/ocr.12138.

5. Lin, Y.H. et al. (2017). LIMCH1 regulates nonmuscle myosin-II activity and suppresses cell migration. *Mol Biol Cell*. E15-04-0218. doi: 10.1091/mbc.E15-04-0218.
6. Wu, T. et al. (2017). Identification of BPIFA1/SPLUNC1 as an epithelium-derived smooth muscle relaxing factor. *Nat Commun*. 8:14118. doi: 10.1038/ncomms14118.
7. Zhu, X. and Jackson, E.K. (2017). RACK1 Regulates Angiotensin II-Induced Contractions of SHR Proliferating Vascular Smooth Muscle Cells. *Am J Physiol Renal Physiol*. 00547.2016. doi: 10.1152/ajprenal.00547.2016
8. Wolfson, B. et al. (2016). High-fat diet promotes mammary gland myofibroblast differentiation through miR-140 downregulation. *Mol. Cell Biol*. doi:10.1128/mcb.00461-16.
9. Zhou, M.W. et al. (2016). Inhibition of collagen synthesis by IWR-1 in normal and keloid-derived skin fibroblasts. *Life Sci*. doi:10.1016/j.lfs.2016.12.003.
10. Duru, N. et al. (2016). Loss of miR-140 is a key risk factor for radiation-induced lung fibrosis through reprogramming fibroblasts and macrophages. *Sci. Rep*. **6**:39572.
11. Loomans, H.A. Esophageal squamous cell carcinoma invasion is inhibited by Activin A in ACVR1B-positive cells. *BMC Cancer* **16**:873.
12. Woeller, C.F. et al. (2016). The aryl hydrocarbon receptor and its ligands inhibit myofibroblast formation and activation: implications for thyroid eye disease. *Am. J. Pathol*. **186**:3189-3202.
13. Greene, W. A. et al. (2016). Secretion profile of induced pluripotent stem cell-derived retinal pigment epithelium during wound healing stem cell-derived RPE profile during wound healing. *Invest Ophthalmol Vis Sci*. **57**:4428-4441.
14. Por, E. D. et al. (2016). Trichostatin A inhibits retinal pigmented epithelium activation in an in vitro model of proliferative vitreoretinopathy. *J Ocul Pharmacol Ther*. doi:10.1089/jop.2016.0038.
15. Jiao, J. et al. (2016). Differentiation defect in neural crest-derived smooth muscle cells in patients with aortopathy associated with bicuspid aortic valves. *EBioMedicine*. doi:10.1016/j.ebiom.2016.06.045.
16. Chen, P.Y. et al. (2016). Smooth muscle FGF/TGF $\beta$  cross talk regulates atherosclerosis progression. *EMBO Mol Med*. doi:10.15252/emmm.201506181.
17. Lazar-Karsten, P. et al. (2016). Generation and characterization of vascular smooth muscle cell lines derived from a patient with a bicuspid aortic valve. *Cells*. doi:10.3390/cells5020019.
18. Ye, Y. et al. (2016). Down-regulation of 14-3-3 Zeta inhibits TGF- $\beta$ 1-induced actomyosin contraction in human trabecular meshwork cells through rhoa signaling pathway 14-3-3 zeta regulates actomyosin contraction in TM cells. *Invest Ophthalmol Vis Sci*. **57**:719-730.
19. Rinella, L. et al. (2016). Extracorporeal shockwaves modulate myofibroblast differentiation of adipose-derived stem cells. *Wound Repair Regen*. doi:10.1111/wrr.12410.
20. Halim, D. et al. (2015). ACTG2 variants impair actin polymerization in sporadic Megacystis Microcolon Intestinal Hypoperistalsis Syndrome. *Hum Mol Genet*. doi:10.1093/hmg/ddv497.
21. Li, H. Y. et al. (2015). Activation of TGF- $\beta$ 1-CD147 positive feedback loop in hepatic stellate cells promotes liver fibrosis. *Sci Rep*. **5**:16552.
22. Ham, S. A. et al. (2015). Ligand-activated PPAR $\delta$  upregulates  $\alpha$ -smooth muscle actin expression in human dermal fibroblasts: a potential role for PPAR $\delta$  in wound healing. *J Dermatol Sci*. doi:10.1016/j.jdermsci.2015.10.005
23. Duru, N. et al. (2015). NRF2/miR-140 signaling confers radioprotection to human lung fibroblasts. *Cancer Lett*. doi:10.1016/j.canlet.2015.08.011.
24. Gutiérrez, J. et al. (2015). RECK-mediated  $\beta$ 1-integrin regulation by TGF- $\beta$ 1 is critical for wound contraction in mice. *PLoS One*. **10**:e0135005.

25. Ratuszny, D. et al. (2015). miR-145 is a promising therapeutic target to prevent cornea scarring. *Hum Gene Ther.* doi:10.1089/hum.2014.151.
26. Sobel, K. et al. (2015). FTY720-P activates sphingosine-1-phosphate receptor 2 and selectively couples to Gα12/13/Rho/ROCK to induce myofibroblast contraction. *MolPharmacol.* doi:10.1124/mol.114.097261.
27. Woeller, C. F. et al. (2015). Salinomycin and other polyether ionophores are a new class of antiscarring agent. *J Biol Chem.* **290**:3563-3575.
28. Luo, Y. et al. (2015). Magnitude-dependent proliferation and contractility modulation of human bladder smooth muscle cells under physiological stretch. *World J Urol.* doi:10.1007/s00345-015-1509-4.
29. Reddy, A. T. et al. (2014). Nitrated fatty acids reverse pulmonary fibrosis by dedifferentiating myofibroblasts and promoting collagen uptake by alveolar macrophages. *FASEB J.* **28**:5299-5310.
30. Rubattu, S. et al. (2014). The C2238/αANP variant is a negative modulator of both viability and function of coronary artery smooth muscle cells. *PLoS One.* **17**:e113108.
31. Hsieh, Y. P. et al. (2014). Arecoline stimulated early growth response-1 production in human buccal fibroblasts: Suppression by epigallocatechin-3-gallate. *Head Neck.* doi:10.1002/hed.23614.
32. Je, Y. J. et al. (2014). Inhibitory role of Id1 on TGF-β-induced collagen expression in human dermal fibroblasts. *Biochem Biophys Res Commun.* **444**:81-85.
33. Herr, M. J. et al. (2014). Tetraspanin CD9 regulates cell contraction and actin arrangement via rhoa in human vascular smooth muscle cells. *PLoS One.* **9**:e106999.
34. Nie, L. et al. (2014). Endothelial-mesenchymal transition in normal human esophageal endothelial cells cocultured with esophageal adenocarcinoma cells: role of IL-1β and TGF-β2. *Am J Physiol Cell Physiol.* **307**:C859-C877.
35. Kotio, K.U. et al. (2011). Implication of microRNAs in Atrial Natriuretic Peptide and Nitric Oxide Signaling in Vascular Smooth Muscle Cells. *Am J Physiol Cell Physiol.* **301**:C929-C937.

## **Warranty**

These products are warranted to perform as described in their labeling and in Cell Biolabs literature when used in accordance with their instructions. THERE ARE NO WARRANTIES THAT EXTEND BEYOND THIS EXPRESSED WARRANTY AND CELL BIOLABS DISCLAIMS ANY IMPLIED WARRANTY OF MERCHANTABILITY OR WARRANTY OF FITNESS FOR PARTICULAR PURPOSE. CELL BIOLABS' sole obligation and purchaser's exclusive remedy for breach of this warranty shall be, at the option of CELL BIOLABS, to repair or replace the products. In no event shall CELL BIOLABS be liable for any proximate, incidental or consequential damages in connection with the products.

## **Contact Information**

Cell Biolabs, Inc.  
 7758 Arjons Drive  
 San Diego, CA 92126  
 Worldwide: +1 858-271-6500  
 USA Toll-Free: 1-888-CBL-0505  
 E-mail: [tech@cellbiolabs.com](mailto:tech@cellbiolabs.com)  
[www.cellbiolabs.com](http://www.cellbiolabs.com)

©2004-2018: Cell Biolabs, Inc. - All rights reserved. No part of these works may be reproduced in any form without permissions in writing.