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Product Manual

# NAD<sup>+</sup>/NADH Assay Kit (Fluorometric)

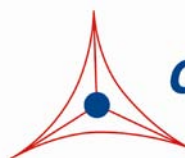
Catalog Number

MET-5030

100 assays

**FOR RESEARCH USE ONLY**  
**Not for use in diagnostic procedures**

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**CELL BIOLABS, INC.**  
*Creating Solutions for Life Science Research*

## **Introduction**

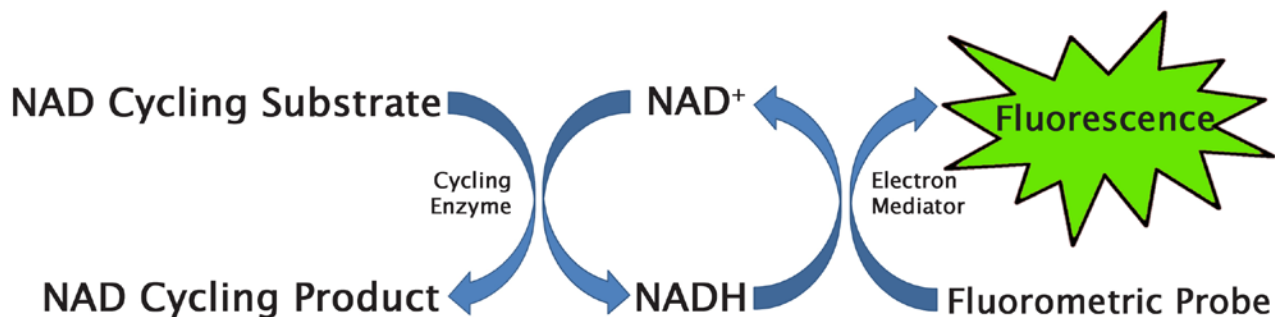
Nicotinamide adenine dinucleotide (NAD) is a complex organic molecule found in all living cells. NAD consists of two nucleotides, adenine and nicotinamide, connected through phosphate groups. NAD exists as an oxidized form ( $\text{NAD}^+$ ) and a reduced form (NADH). In cells,  $\text{NAD}^+$  acts as an electron acceptor, becoming reduced from other molecules to form NADH. The resulting NADH can then act as a reducing agent, donating electrons. An example of this occurs when reduced compounds, such as glucose and fatty acids, become oxidized and transfer energy via electrons to  $\text{NAD}^+$  to form NADH. This electron transfer chemistry is observed in glycolysis and the citric acid cycle. In eukaryotic cells, NADH electrons made in the cytoplasm are transported to mitochondrial  $\text{NAD}^+$  by mitochondrial shuttles such as the malate-aspartate shuttle. The mitochondrial NADH is then oxidized by the electron transport chain that pumps protons across a membrane, generating ATP through a process known as oxidative phosphorylation.

In addition to electron transfer,  $\text{NAD}^+$  and NADH are also used as enzyme substrates to add or remove posttranslational modifications from proteins, as in the process of ADP ribosylation.  $\text{NAD}^+$  is also consumed by NAD-dependent deacetylases known as sirtuins. These enzymes can transfer an acetyl group from their substrate protein to the ADP-ribose of NAD, which then cleaves the coenzyme and releases nicotinamide and O-acetyl-ADP-ribose. Non-eukaryotic DNA ligases, which join two DNA ends, also operate in an  $\text{NAD}^+$  dependent fashion by using  $\text{NAD}^+$  as a substrate to donate an adenosine monophosphate (AMP) moiety to one of the 5' phosphate DNA ends. The resulting intermediate is then attacked by the 3' hydroxyl group of the other DNA end, forming the final phosphodiester bond.  $\text{NAD}^+$  has also been identified as an extracellular signaling molecule.  $\text{NAD}^+$  is released from neurons in the large intestine, blood vessels, urinary bladder, neurosecretory cells and from brain synaptosomes.  $\text{NAD}^+$  is therefore a novel neurotransmitter that transmits a signal from nerves to effector cells.

Cell Biolabs'  $\text{NAD}^+$ /NADH Assay Kit is a simple fluourometric assay that measures  $\text{NAD}^+$  and NADH present in biological samples such as cell lysates or tissue extracts in a 96-well microtiter plate format. The kit is specific for  $\text{NAD}^+$ , NADH, and their ratio. The kit will not detect  $\text{NADP}^+$  or  $\text{NADPH}$ . Each kit provides sufficient reagents to perform up to 100 assays, including blanks,  $\text{NAD}^+$  standards and unknown samples. The total  $\text{NAD}^+$ /NADH concentrations of unknown samples are determined by comparison with a known  $\text{NAD}^+$  standard. Determination of both  $\text{NAD}^+$  and NADH requires two separate samples for quantification.  $\text{NAD}^+$  and NADH do not need to be purified from samples, but rather can be extracted individually with a simple acid or base treatment prior to performing the assay. The kit has a detection sensitivity limit of approximately 0.8 nM  $\text{NAD}^+$ .

## **Assay Principle**

Cell Biolabs'  $\text{NAD}^+$  /NADH Assay Kit is a convenient quantitative tool that measures  $\text{NAD}^+$  and NADH within biological samples. The assay is based on an enzymatic cycling reaction in which  $\text{NAD}^+$  is reduced to NADH. NADH reacts with a fluorometric probe that produces a product which can be measured with a standard 96-well fluorometric plate reader. The intensity of the product fluorescence is proportional to the  $\text{NAD}^+$  and NADH within a sample. A simple acid or base treatment will differentiate NADH from  $\text{NAD}^+$  within a sample. Samples and standards are incubated for 1-2 hours and then read at  $\lambda_{\text{ex}} = 530\text{-}570 \text{ nm}$  /  $\lambda_{\text{em}} = 590\text{-}600 \text{ nm}$  (Figure 1). Samples are compared to a known concentration of  $\text{NAD}^+$  standard within the 96-well microtiter plate format.



**Figure 1. NAD<sup>+</sup>/NADH Cycling Assay Principle.**

### **Related Products**

1. MET-5014: NAD<sup>+</sup>/NADH Assay Kit
2. MET-5018: NADP<sup>+</sup>/NADPH Assay Kit
3. MET-5031: NADP<sup>+</sup>/NADPH Assay Kit (Fluorometric)
4. STA-312: Total Glutathione Assay Kit
5. STA-384: Total Cholesterol Assay Kit (Colorimetric)
6. STA-618: Free Fatty Acid Assay Kit (Colorimetric)
7. STA-631: Total Bile Acid Assay Kit (Colorimetric)
8. STA-680: Glucose Assay Kit (Colorimetric)
9. STA-812: Glutathione Reductase Assay Kit
10. XPX-5006: Monoamine Oxidase Assay Kit (Colorimetric)

### **Kit Components**

#### **Box 1 (shipped at room temperature)**

1. Fluorometric Probe (40X) (Part No. 50311C): One 150 µL amber tube
2. NAD Cycling Substrate (Part No. 50142B): One 200 µL tube
3. Assay Buffer (Part No. 50184B): One 25 mL bottle
4. Extraction Buffer (10X) (Part No. 50185B): One 10 mL bottle
5. Electron Mediator (500X) (Part No. 50312C): One 10 µL amber tube

#### **Box 2 (shipped on blue ice packs)**

1. NAD Cycling Enzyme (Part No. 50144D): One 50 µL amber tube
2. NAD<sup>+</sup> Standard (Part No. 50141D): One 50 µL amber tube of a 20 mM NAD<sup>+</sup> solution

## **Materials Not Supplied**

1. Distilled or deionized water
2. 0.1 N NaOH
3. 0.1 N HCl
4. 10 kDa molecular weight cutoff (MWCO) centrifuge spin filter (e.g. Amicon Ultra 0.5mL)
5. 1X PBS
6. 10  $\mu$ L to 1000  $\mu$ L adjustable single channel micropipettes with disposable tips
7. 50  $\mu$ L to 300  $\mu$ L adjustable multichannel micropipette with disposable tips
8. Standard 96-well fluorescence microtiter plate and/or fluorescence cell culture microplate
9. Multichannel micropipette reservoir
10. Fluorescence microplate reader capable of reading excitation in the 530-570 nm range and emission in the 590-600 nm range.

## **Storage**

Upon receipt, store the NAD Cycling Enzyme and NAD<sup>+</sup> Standard at -80°C. Store the Fluorometric Probe and Electron Mediator at -20°C. Store the remaining components at 4°C.

## **Preparation of Reagents**

- 1X Extraction Buffer: Dilute the stock 10X Extraction Buffer 1:10 with deionized water for a 1X solution. Stir or vortex to homogeneity.
- NAD Cycling Reagent: Prepare an NAD Cycling Reagent for the number of assays being tested and just before use. Prepare by diluting the NAD Cycling Substrate 1:25, NAD Cycling Enzyme 1:100, Electron Mediator 1:500, and Fluorometric Probe 1:40 in 1X Assay Buffer. (eg. For 100 assays, combine 200  $\mu$ L NAD Cycling Substrate, 50  $\mu$ L NAD Cycling Enzyme, 10  $\mu$ L Electron Mediator, and 125  $\mu$ L Fluorometric Probe to 4.615 mL of 1X Assay Buffer for a 5 mL total solution). When preparing, combine the NAD Cycling Substrate, NAD Cycling Enzyme, and Electron Mediator together first and mix prior to adding the Fluorometric Probe. Upon adding the probe, protect the solution from light to prevent the probe from oxidizing. Use the NAD Cycling Reagent immediately.

## **Preparation of Samples**

These preparation protocols are intended as a guide for preparing unknown samples. The user may need to adjust the sample treatment accordingly. It is highly recommended that all samples should be assayed immediately upon preparation or stored for up to 1 month at -80°C. A trial assay with a representative test sample should be performed to determine the sample compatibility with the dynamic range of the standard curve. High levels of interfering substances may cause variations in results. Samples may be diluted in deionized water as necessary before testing. Run proper controls and account for any sample dilutions. Always run a standard curve with samples.

- Tissue homogenates: Sonicate or homogenize 100 mg tissue sample in 0.5 mL cold 1X Extraction Buffer. Centrifuge at 14,000 rpm for 5 minutes at 4°C to remove insoluble material. Filter the solution with a 10kDa spin filter to deproteinate the sample. Collect flow through. Perform dilutions in cold deionized water. Sample may be tested immediately for total NAD<sup>+</sup>/NADH quantification or extracted with acid or base to separate the cofactors. Store unused samples at -80°C for up to 1 month.
- Cell lysates: Culture cells until confluent and harvest. Centrifuge and wash cell pellet with 1X PBS. Centrifuge to pellet cells and remove wash. Resuspend cells at 1-5 x 10<sup>6</sup> cells/mL in 0.5 mL 1X Extraction Buffer. Homogenize or sonicate the cells on ice. Centrifuge at 14,000 rpm for 5 minutes 4°C to remove debris. Filter the solution with a 10kDa spin filter to deproteinate the sample. Collect flow through. Perform dilutions in cold deionized water. Sample may be tested immediately for total NAD<sup>+</sup>/NADH quantification or extracted with acid or base to separate the cofactors. Store unused samples at -80°C for up to 1 month.

*Note: Enzymes in tissue and cell sample lysates may deplete NADH rapidly and affect results. Samples should be deproteinized before extracting the cofactors or using within the assay. A spin filter with a 10 kDa cutoff is recommended for efficient and clean separation.*

#### **NADH Extraction Procedure:**

To measure NADH and destroy NAD<sup>+</sup>, add 25 µL of sample to a microcentrifuge tube. Add 5 µL of 0.1 N NaOH and mix thoroughly. Incubate the tube at 80°C for 60 minutes and protected from light. Centrifuge the tube to pool all sample solution. Add 20 µL of 1X Assay Buffer to shift the pH of the sample back to neutral. Vortex to mix and centrifuge to pool sample. Sample pH should be between 6.0 and 8.0; if not, neutralize accordingly with acid or base. Keep sample on ice until assaying.

#### **NAD<sup>+</sup> Extraction Procedure:**

To measure NAD<sup>+</sup> and destroy NADH, add 25 µL of sample to a microcentrifuge tube. Add 5 µL of 0.1 N HCl and mix thoroughly. Incubate the tube at 80°C for 60 minutes and protected from light. Centrifuge the tube to pool all sample solution. Add 20 µL of 1X Assay Buffer to shift the pH of the sample back to neutral. Vortex to mix and centrifuge to pool sample. Sample pH should be between 6.0 and 8.0; if not, neutralize accordingly with acid or base. Keep sample on ice until assaying.

#### *Notes:*

- *Samples should be close to neutral pH before harvesting. Samples with extremely high or low pH values could fail to yield reliable results.*
- *If testing both total NAD<sup>+</sup>/NADH and individual cofactors, dilute the total NAD<sup>+</sup>/NADH samples 1:2 with Assay buffer to maintain sample dilution consistency.*
- *Avoid samples containing SH groups like DTT, β-mercaptoethanol, or reduced glutathione.*

#### **Preparation of Standard Curve**

Prepare NAD<sup>+</sup> standards immediately before use. First, dilute the stock 20 mM NAD<sup>+</sup> Standard solution 1:1000 in deionized water to yield a 20 µM NAD<sup>+</sup> solution (e.g. add 5 µL of the stock 20 mM NAD<sup>+</sup> Standard to 4.995 mL of deionized water). Further dilute the 20 µM NAD<sup>+</sup> solution 1:100 in deionized water to yield a 0.2 µM (200 nM) solution (e.g. add 10 µL of the 20 µM NAD<sup>+</sup> solution to

990  $\mu\text{L}$  of deionized water). Use this 200 nM solution to prepare a series of  $\text{NAD}^+$  standards according to Table 1 below. Do not store standard solutions.

Standard Tubes	200 nM $\text{NAD}^+$ Solution ( $\mu\text{L}$ )	1X Extraction Buffer ( $\mu\text{L}$ )	$\text{NAD}^+$ (nM)	$\text{NAD}^+$ (pmol/well)
1	250	250	100	5
2	250 of Tube #1	250	50	2.5
3	250 of Tube #2	250	25	1.25
4	250 of Tube #3	250	12.5	0.625
5	250 of Tube #4	250	6.25	0.313
6	250 of Tube #5	250	3.13	0.156
7	250 of Tube #6	250	1.56	0.078
8	250 of Tube #7	250	0.78	0.039
9	0	250	0	0

**Table 1. Preparation of  $\text{NAD}^+$  Standards**

### **Assay Protocol**

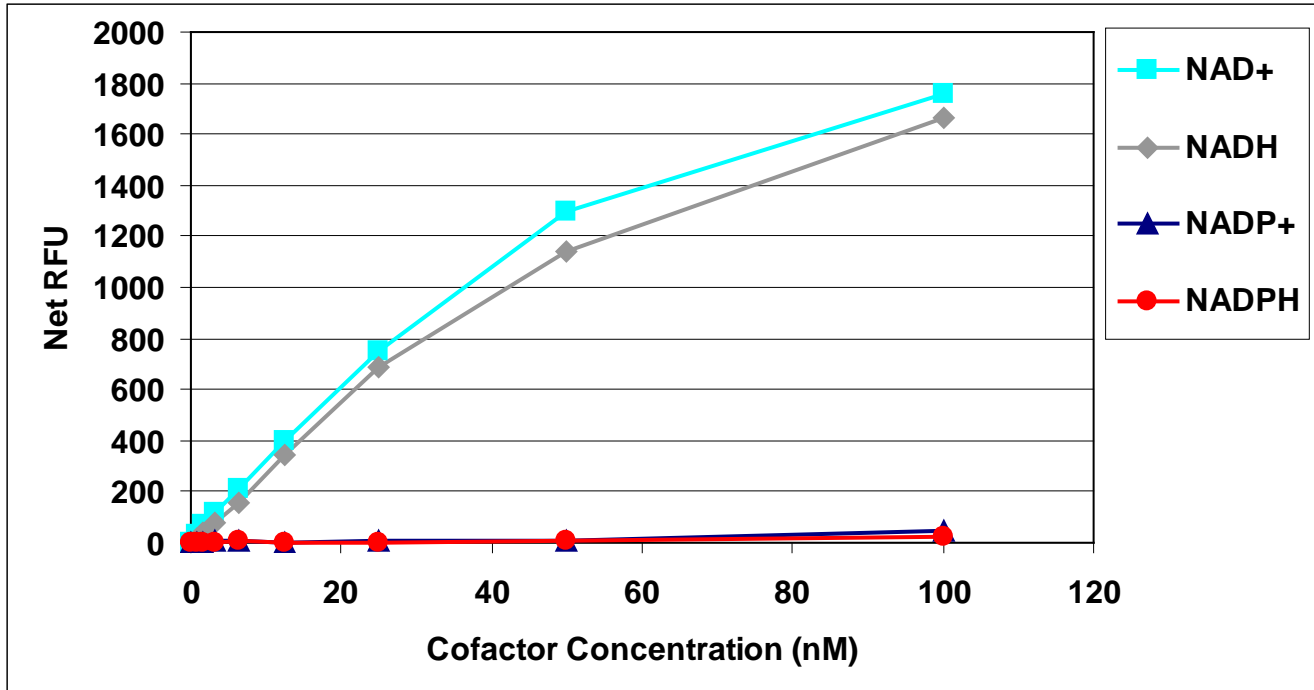
1. Prepare and mix all reagents thoroughly before use. Each sample, including unknowns and standards, should be assayed in duplicate or triplicate.
2. Add 50  $\mu\text{L}$  of each  $\text{NAD}^+$  standard or unknown sample into wells of a 96-well microtiter plate.
3. Add 50  $\mu\text{L}$  of NAD Cycling Reagent to each well. Mix the well contents thoroughly and incubate for 1-2 hours at room temperature protected from light.

*Note: This assay is continuous (not terminated) and therefore may be measured at multiple time points to follow the reaction kinetics.*

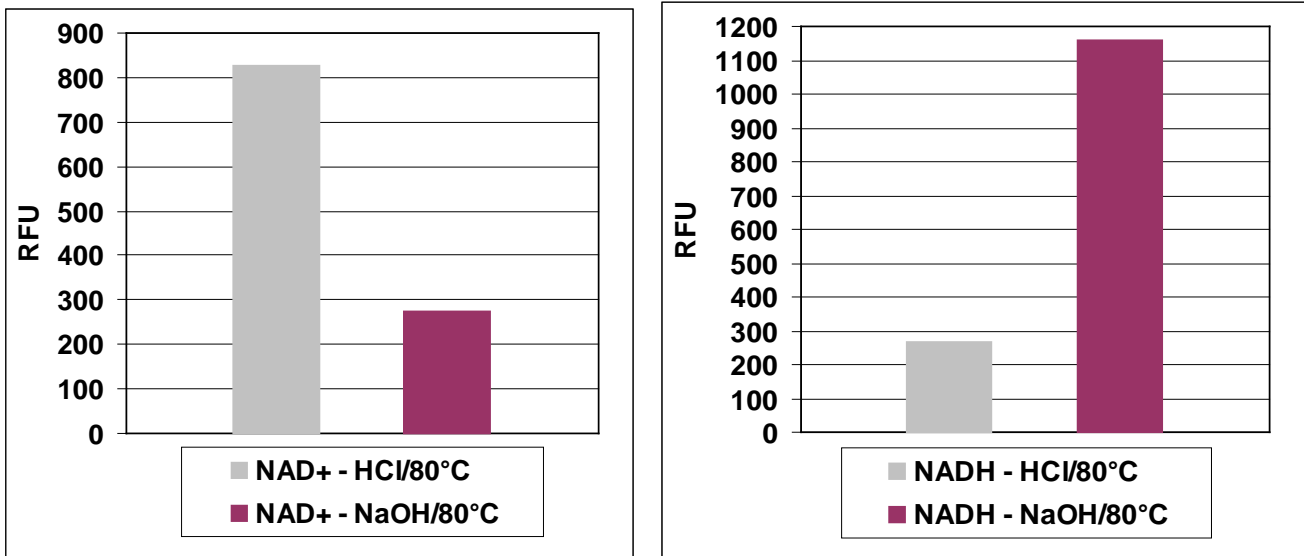
4. Read the plate with a fluorescence microplate reader equipped for excitation in the 530-570 nm range and for emission in the 590-600 nm range.
5. Calculate the concentration of  $\text{NAD}^+/\text{NADH}$  within samples by comparing the sample RFU to the standard curve.

### **Example of Results**

The following figures demonstrate typical  $\text{NAD}^+/\text{NADH}$  Assay Kit results. One should use the data below for reference only. This data should not be used to interpret or calculate actual sample results.



**Figure 2: NAD<sup>+</sup> Standard Curve and Specificity of Assay for NAD<sup>+</sup> and NADH.** NAD<sup>+</sup>, NADH, NADP<sup>+</sup>, and NADPH were tested in the NAD<sup>+</sup>/NADH Assay Kit.



**Figure 3: NAD<sup>+</sup>/NADH Detection.** NAD<sup>+</sup> and NADH were both tested at 100 nM with the extraction procedure. NAD<sup>+</sup> or NADH were incubated for 60 minutes at 80°C with 0.1N HCl or 0.1N NaOH.

### Calculation of Results

1. Calculate the average fluorescence values for every standard, control, and sample. Subtract the average zero standard value from itself and all standard and sample values. This is the corrected

background fluorescence. If sample background control value is high, subtract the sample background control value from the sample reading.

2. Plot the corrected fluorescence for the NAD<sup>+</sup> standards against the final concentration of the standards from Table 1 to determine the best slope (nM<sup>-1</sup>). See Figure 2 for an example standard curve.
3. Since all NAD<sup>+</sup> is converted to NADH by the Cycling Reagent, use the standard curve to determine the total NAD<sup>+</sup>/NADH concentration in pmoles within the sample. Determine the total concentration of the samples with the equation obtained from the linear regression analysis of the standard curve. Substitute the corrected fluorescence values for each sample. Remember to account for dilution factors.

$$\text{Total NAD}^+/\text{NADH} = \left[ \frac{\text{Sample corrected fluorescence}}{\text{Slope}} \right] \times \text{Sample dilution}$$

## References

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## Warranty

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