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Product Manual

# OxiSelect™ Oxidative DNA Damage Quantitation Kit (AP Sites)

Catalog Number

STA-324

50 assays

**FOR RESEARCH USE ONLY**  
Not for use in diagnostic procedures

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**CELL BIOLABS, INC.**  
*Creating Solutions for Life Science Research*

## **Introduction**

Free radicals and other reactive species are constantly generated *in vivo* and cause oxidative damage to biomolecules, a process held in check only by the existence of multiple antioxidant and repair systems as well as the replacement of damaged lipids and proteins. DNA is probably the most biologically significant target of oxidative attack, and it is widely thought that continuous oxidative damage to DNA is a significant contributor to the age-related development of the major cancers, such as those of the colon, breast, rectum, and prostate. Among numerous types of oxidative DNA damage, apurinic/aprimidinic (AP or abasic) site is one of the prevalent lesions of oxidative DNA damage. Abasic sites arise in DNA at a significant rate by spontaneous base loss as in depurination, by DNA oxidation, or by the action of DNA glycosylases. Estimates of the number of abasic sites generated per mammalian cell run as high as 50,000 to 200,000 per day. Unrepaired abasic sites inhibit topoisomerases, replication, and transcription and can be mutagenic because of bypass synthesis on nontemplated DNA.

The OxiSelect™ Oxidative DNA Damage Quantitation Kit (AP sites) uses an Aldehyde Reactive Probe (ARP) to react specifically with an aldehyde group on the open ring form of AP sites. This allows for the AP sites to be tagged with biotin which is later detected with Streptavidin-Enzyme conjugate. The quantity of AP sites in unknown DNA sample is determined by comparing its absorbance with a standard curve generated from the provided DNA standard containing predetermined AP sites. The kit has a detection sensitivity range of 4 to 40 AP sites per  $1 \times 10^5$  bp. Each kit provides sufficient reagents to perform up to 50 assays for unknown samples, excluding the standard curve.

## **Related Products**

1. STA-320: OxiSelect™ Oxidative DNA Damage ELISA Kit (8-OHdG Quantitation)
2. STA-321: OxiSelect™ DNA Double-Strand Break (DSB) Staining Kit
3. STA-325: OxiSelect™ Oxidative RNA Damage ELISA Kit (8-OHG Quantitation)
4. STA-350: OxiSelect™ Comet Assay Kit (3-Well Slides), 15 Assays
5. STA-351: OxiSelect™ Comet Assay Kit (3-Well Slides), 75 Assays
6. STA-352: OxiSelect™ Comet Assay Slides (3-Well), 5 Slides
7. STA-353: OxiSelect™ Comet Assay Slides (3-Well), 25 Slides
8. STA-355: OxiSelect™ 96-Well Comet Assay Kit
9. STA-356: OxiSelect™ 96-Well Comet Assay Slide

## **Kit Components**

1. Glycogen Solution (Part No. 232401): One 100  $\mu$ L vial of 10 mg/mL glycogen.
2. Sodium Acetate Solution (Part No. 232402): One 1.0 mL vial of 3M Sodium Acetate, pH 5.5.
3. ARP Solution (Part No. 232403): One 250  $\mu$ L vial of 10 mM ARP.
4. DNA High-Binding Plate (Part No. 232404): One 96-well strip plate.
5. DNA Binding Solution (Part No. 232405): One 6 mL bottle.

6. 10X Wash Buffer (Part No. 232406): One 30 mL bottle.
7. Streptavidin-Enzyme Conjugate (Part No. 310803): One 20  $\mu$ L vial.
8. Substrate Solution (Part No. 310807): One 12 mL amber bottle.
9. Stop Solution (Part No. 310808): One 12 mL bottle.
10. Reduced DNA Standard (Part No. 232407): One 1.0 mL vial of 6  $\mu$ g/mL fully reduced in TE Buffer (0 ARP/100,000 bp).
11. ARP-DNA Standard (Part No. 232408): One 400  $\mu$ L vial of 6  $\mu$ g/mL ARP-DNA in TE Buffer (40 ARP/100,000 bp).

### **Materials Not Supplied**

1. DNA samples from cell or tissue for measuring DNA damage
2. TE Buffer: 10 mM Tris, pH 7.5, 1 mM EDTA
3. 100% and 70% Ethanol
4. 10  $\mu$ L to 1000  $\mu$ L adjustable single channel micropipettes with disposable tips
5. 50  $\mu$ L to 300  $\mu$ L adjustable multichannel micropipette with disposable tips
6. 37°C Incubator
7. Multichannel micropipette reservoir
8. Microplate reader capable of reading at 450 nm (620 nm as optional reference wave length)

### **Storage**

Upon receipt, aliquot and store both the Reduced DNA and ARP-DNA Standards at -20°C to avoid multiple freeze/thaw cycles. Store all other components at 4°C.

### **Preparation of Reagents**

- 1X Wash Buffer: Dilute the 10X Wash Buffer Concentrate to 1X with deionized water. Stir to homogeneity.
- Streptavidin-Enzyme Conjugate: Immediately before use, dilute the Streptavidin-Enzyme Conjugate 1:1000 with 1X Wash Buffer. Do not store diluted solutions.

### **Preparation of Standard Curve**

Prepare a dilution series of ARP-DNA standards in the concentration range of 0 – 40 ARP/100,000 bp according to Table 1.

Tubes	ARP-DNA Standard (μL)	Reduced DNA Standard (μL)	TE Buffer (μL)	Total Volume (μL)	DNA Concentration (μg/mL)	AP Sites per 100,000 bp
1	20	0	100	120	1	40
2	16	4	100	120	1	32
3	12	8	100	120	1	24
4	8	12	100	120	1	16
5	4	16	100	120	1	8
6	2	18	100	120	1	4
7	1	19	100	120	1	2
8	0	20	100	120	1	0

**Table 1. Preparation of ARP-DNA Standards**

## **Assay Protocol**

### **I. ARP Reaction**

1. Isolate genomic DNA with desired method and dissolve the genomic DNA in TE buffer. Dilute the genomic DNA with TE buffer to 100 μg/mL.

*Note: During DNA extraction, avoid heating the DNA solution, or any procedure will introduce AP sites. We recommend using DNAZOL reagent to extract DNA and dissolve DNA in TE buffer.*

2. Mix 5 μL of purified genomic DNA (100 μg/mL) with 5 μL of ARP solution in a microcentrifuge tube and incubate 1 hr at 37°C.
3. Add 90 μL of TE buffer and 1 μL of Glycogen Solution to each tube and mix well.
4. Add 10 μL of Sodium Acetate Solution to each tube, mix well.
5. Add 300 μL of absolute ethanol to each tube and mix well and incubate at -20°C for 30 minutes.
6. Centrifuge for 10-20 minutes at 14,000 g and carefully wash the pellet three times with 70% ethanol.
7. Dissolve the DNA pellet in 10-50 μL of TE buffer and determine the DNA concentration with desired method. ARP-derived DNA can be stored at -20°C for up to one year.

*Note: It is important that the ARP-derived DNA concentration is determined precisely for the accurate measurement of AP sites. We recommend using Invitrogen's Quanti-iT™ DNA assay kit to measure DNA concentration.*

## II. Determination of AP sites in DNA:

1. Dilute the ARP-derived DNA sample to 1  $\mu\text{g}/\text{mL}$  with TE buffer.
2. Add 50  $\mu\text{L}$  of ARP-derived DNA sample or each dilution of the prepared ARP-DNA standards to the DNA High-binding plate. Add 50  $\mu\text{L}$  of DNA Binding Solution to each well. Mix well by pipetting and incubate at room temperature for 2 hrs or overnight on an orbital shaker. Each sample including unknown and standard should be assayed in duplicate.
3. Wash microwell strips 3 times with 250  $\mu\text{L}$  1X Wash Buffer per well with thorough aspiration between each wash. After the last wash, empty wells and tap microwell strips on absorbent pad or paper towel to remove excess 1X Wash Buffer.
4. Add 100  $\mu\text{L}$  of diluted Streptavidin-Enzyme Conjugate to each well and incubate at 37°C for 1 hr.
5. Wash microwell strips 3 times with 250  $\mu\text{L}$  1X Wash Buffer per well with thorough aspiration between each wash. After the last wash, empty wells and tap microwell strips on absorbent pad or paper towel to remove excess 1X Wash Buffer.
6. Warm Substrate Solution to room temperature. Add 100  $\mu\text{L}$  of Substrate Solution to each well, including the blank wells. Incubate at room temperature for 5 to 20 minutes on an orbital shaker.
7. Stop the enzymatic reaction by adding 100  $\mu\text{L}$  of Stop Solution into each well, including the blank wells. Results should be read immediately (color will fade over time).
8. Read absorbance of each microwell on a spectrophotometer using 450 nm as the primary wave length.

## Example of Results

The following figures demonstrate typical Oxidative DNA Damage Quantitation results. One should use the data below for reference only. This data should not be used to interpret actual results.

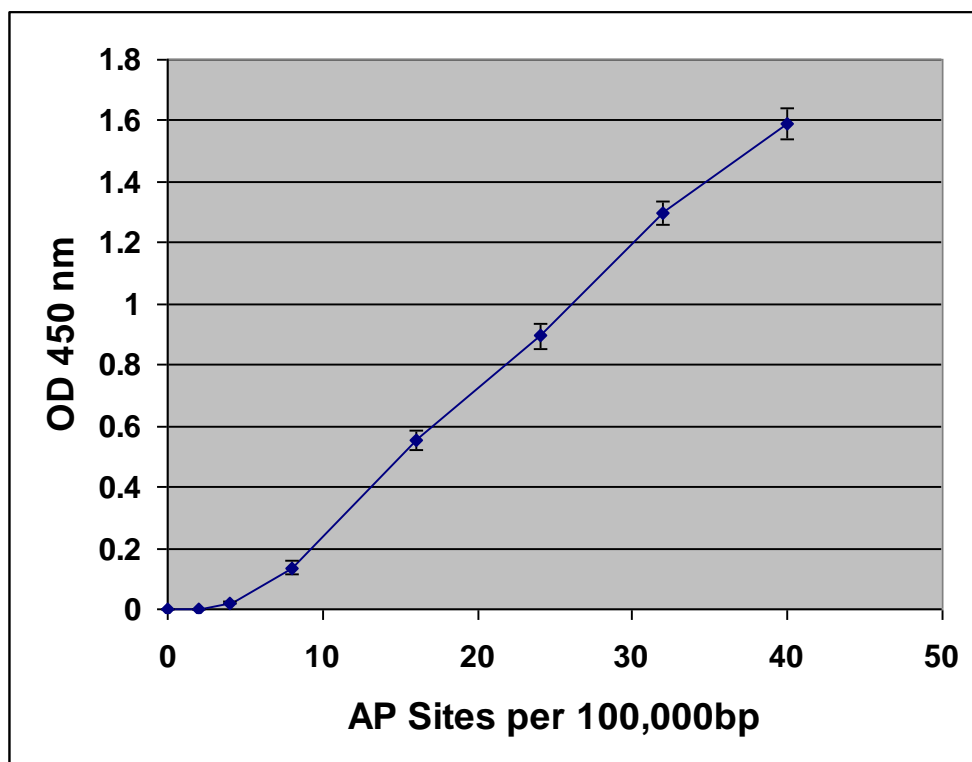


Figure 1: ARP-DNA Standard Curve.

## References

1. Croteau D L, Bohr V A. (1997) *J Biol Chem.* **272**:25409–25412.
2. Lindahl T. (1993) *Nature* **362**:709–715.
3. Kubo K, Ide H, Wallace S S, Kow Y W. (1992) *Biochemistry* **31**:3703–3708.

## Recent Product Citations

1. Sfrikakis, P.P. et al. (2022). Microvasculopathy-Related Hemorrhagic Tissue Deposition of Iron May Contribute to Fibrosis in Systemic Sclerosis: Hypothesis-Generating Insights from the Literature and Preliminary Findings. *Life (Basel)*. **12**(3):430. doi: 10.3390/life12030430.
2. Talluri, S. et al. (2021). Dysregulated APOBEC3G causes DNA damage and promotes genomic instability in multiple myeloma. *Blood Cancer J.* **11**(10):166. doi: 10.1038/s41408-021-00554-9.
3. Haider, N. et al. (2021). Signaling defects associated with insulin resistance in non-diabetic and diabetic individuals and modification by sex. *J Clin Invest.* doi: 10.1172/JCI151818.
4. Ognik, K. et al. (2021). The immune status, oxidative and epigenetic changes in tissues of turkeys fed diets with different ratios of arginine and lysine. *Sci Rep.* **11**(1):15975. doi: 10.1038/s41598-021-95529-y.

5. Kim, J.H. et al. (2021). Nordihydroguaiaretic Acid as a Novel Substrate and Inhibitor of Catechol O-Methyltransferase Modulates 4-Hydroxyestradiol-Induced Cyto- and Genotoxicity in MCF-7 Cells. *Molecules*. **26**(7):2060. doi: 10.3390/molecules26072060.
6. Psyri, A. et al. (2021). The DNA damage response network in the treatment of head and neck squamous cell carcinoma. *ESMO Open*. **6**(2):100075. doi: 10.1016/j.esmoop.2021.100075.
7. Dettleff, P. et al. (2020). Physiological and molecular responses to thermal stress in red cusk-eel (*Genypterus chilensis*) juveniles reveals atrophy and oxidative damage in skeletal muscle. *J Therm Biol*. doi: 10.1016/j.jtherbio.2020.102750.
8. Ognik, K. et al. (2020). The effect of different dietary ratios of lysine and arginine in diets with high or low methionine levels on oxidative and epigenetic DNA damage, the gene expression of tight junction proteins and selected metabolic parameters in *Clostridium perfringens*-challenged turkeys. *Vet Res*. **51**(1):50. doi: 10.1186/s13567-020-00776-y.
9. Huo, X. et al. (2020). In Barrett's Epithelial Cells, Weakly Acidic Bile Salt Solutions Cause Oxidative DNA Damage with Response and Repair Mediated by p38. *Am J Physiol Gastrointest Liver Physiol*. doi: 10.1152/ajpgi.00329.2019.
10. Sherwood, T.A. et al. (2019). Nonlethal Biomarkers of Oxidative Stress in Oiled Sediment Exposed Southern Flounder (*Paralichthys lethostigma*): Utility for Field-Base Monitoring Exposure and Potential Recovery. *Environ Sci Technol*. **53**(24):14734-14743. doi: 10.1021/acs.est.9b05930.
11. Rivas-Aravena, A. et al. (2019). Transcriptomic response of rainbow trout (*Oncorhynchus mykiss*) skeletal muscle to *Flavobacterium psychrophilum*. *Comparative Biochemistry and Physiology Part D: Genomics and Proteomics*. 100596. doi: 10.1016/j.cbd.2019.100596.
12. Thai, S.F. et al. (2019). Differential Effects of Nano TiO<sub>2</sub> and CeO<sub>2</sub> on Normal Human Lung Epithelial Cells In Vitro. *J Nanosci Nanotechnol*. **19**(11):6907-6923. doi: 10.1166/jnn.2019.16737.
13. Norambuena, J. et al. (2019). Superoxide Dismutase and Pseudocatalase Increase Tolerance to Hg(II) in *Thermus thermophilus* HB27 by Maintaining the Reduced Bacillithiol Pool. *MBio*. **10**(2). pii: e00183-19. doi: 10.1128/mBio.00183-19.
14. Souliotis, V.L. et al. (2019). DNA damage accumulation, defective chromatin organization and deficient DNA repair capacity in patients with rheumatoid arthritis. *Clin Immunol*. **203**:28-36. doi: 10.1016/j.clim.2019.03.009.
15. Patchsung, M. et al. (2018). Alu siRNA to increase Alu element methylation and prevent DNA damage. *Epigenomics*. **10**(2):175-185. doi: 10.2217/epi-2017-0096.
16. Mishra, A. et al. (2018). Oxidative Stress-Mediated Overexpression of Uracil DNA Glycosylase in *Leishmania donovani* Confers Tolerance against Antileishmanial Drugs. *Oxid Med Cell Longev*. **2018**:4074357. doi: 10.1155/2018/4074357.
17. Thakur, S. et al. (2017). APE1 modulates cellular responses to organophosphate pesticide-induced oxidative damage in non-small cell lung carcinoma A549 cells. *Molecular and Cellular Biochemistry*. **441**(1-2): 201-216.
18. Mullick, M. et al. (2017). d-Alanine 2, Leucine 5 Enkephaline (DADLE)-mediated DOR activation augments human hUCB-BFs viability subjected to oxidative stress via attenuation of the UPR. *Stem Cell Res*. **22**:20-28. doi: 10.1016/j.scr.2017.05.009.
19. Periyasamy, M. et al (2017). p53 controls expression of the DNA deaminase APOBEC3B to limit its potential mutagenic activity in cancer cells. *Nucleic Acids Research*, gkx721.
20. Stasiolek, M. et al. (2017). The molecular effect of diagnostic absorbed doses from <sup>131</sup>I on papillary thyroid cancer cells in vitro. *Molecules* doi:10.3390/molecules22060993.

21. Sapoznik, S. et al. (2016). Activation-induced cytidine deaminase links ovulation-induced inflammation and serous carcinogenesis. *Neoplasia*. **18**:90-99.
22. Garama, D. J. et al. (2015). A synthetic lethal interaction between glutathione synthesis and mitochondrial reactive oxygen species provides a tumor specific vulnerability dependent on STAT3. *Mol Cell Biol*. doi:10.1128/MCB.00541-15.
23. Guzmán-Guillén, R. et al. (2015). Beneficial effects of vitamin E supplementation against the oxidative stress on Cylindrospermopsin-exposed tilapia (*Oreochromis niloticus*). *Toxicol*. **104**:34-42.
24. Ferreira, E. et al. (2015). Glyceraldehyde-3-Phosphate Dehydrogenase is required for efficient repair of cytotoxic DNA lesions in *Escherichia coli*. *Int J Biochem Cell Biol*. **60**:202-212.
25. Zhao, K. et al. (2014). S-sulfhydration of MEK1 leads to PARP-1 activation and DNA damage repair. *EMBO Rep*. **15**:792-800.
26. Mohammad, M. K. et al. (2014). Watermelon (*Citrullus lanatus* (Thunb.) Matsum. and Nakai) juice modulates oxidative damage induced by low dose X-ray in mice. *Biomed Res Int*. **2014**:512834.
27. Zafiropoulos, A. et al. (2014). Cardiotoxicity in rabbits after a low-level exposure to diazinon, propoxur, and chlorpyrifos. *Hum Exp Toxicol*. **33**:1241-1252.
28. Messaoudi, N. et al. (2013). Global stress response in a prokaryotic model of DJ-1-associated Parkinsonism. *J.Bacteriol*. **195**:1167-1178.
29. Zaika, E. et al. (2011). p73 protein regulates DNA damage repair. *FASEB J*. **25**:4406-4414.

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