
Product Manual

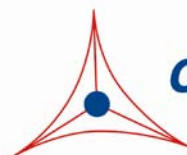
HDL-Cholesterol Assay Kit

Catalog Number

STA-394

96 assays

FOR RESEARCH USE ONLY
Not for use in diagnostic procedures



CELL BIOLABS, INC.
Creating Solutions for Life Science Research

Introduction

Cholesterol is a lipid sterol that is produced in and transported throughout the bloodstream in eukaryotes. Cholesterol is a critical compound used in the structure of cell membranes, hormones, and cell signaling. It is an essential component of animal cell structure in order to maintain permeability and fluidity. Cholesterol is a precursor for steroid hormones, including the adrenal gland hormones cortisol and aldosterone, sex hormones progesterone, estrogens, and testosterone, as well as bile acids, and vitamin D. Cholesterol is transported around the body within lipoproteins, which are submicroscopic particles composed of lipid and protein held together by noncovalent forces. Their general structure is that of a putative spheroidal microemulsion formed from an outer layer of phospholipids, unesterified cholesterol, and proteins, with a core of neutral lipids, predominately cholesteryl esters and triacylglycerols (TAG). Lipoprotein's main function is to transport these lipids around the body in the blood.

Lipoprotein particles have hydrophilic groups of phospholipids, cholesterol, and apoproteins directed outward. Such characteristics make them soluble in the salt water-based blood pool. Triglyceride-fats and cholesterol esters are carried internally, shielded from the water by the phospholipid monolayer and the apoproteins. The interaction of the proteins forming the surface of the particles with enzymes in the blood, with each other, and with specific proteins on the surfaces of cells determine whether triglycerides and cholesterol will be added to or removed from the lipoprotein transport particles. Lipoproteins have cell-specific signals that direct the lipids they transport to certain tissues. For this reason, lipoproteins exist in different forms within the blood based on their density. These include chylomicrons, very-low density lipoproteins (VLDLs), intermediate-density lipoproteins (IDLs), low-density lipoproteins (LDLs), and high-density lipoproteins (HDLs). The higher the lipid content in a lipoprotein, the less dense it is. Cholesterol exists within a lipoprotein as a free alcohol and as a fatty cholesteryl ester, which is the predominant form of cholesterol transport and storage.

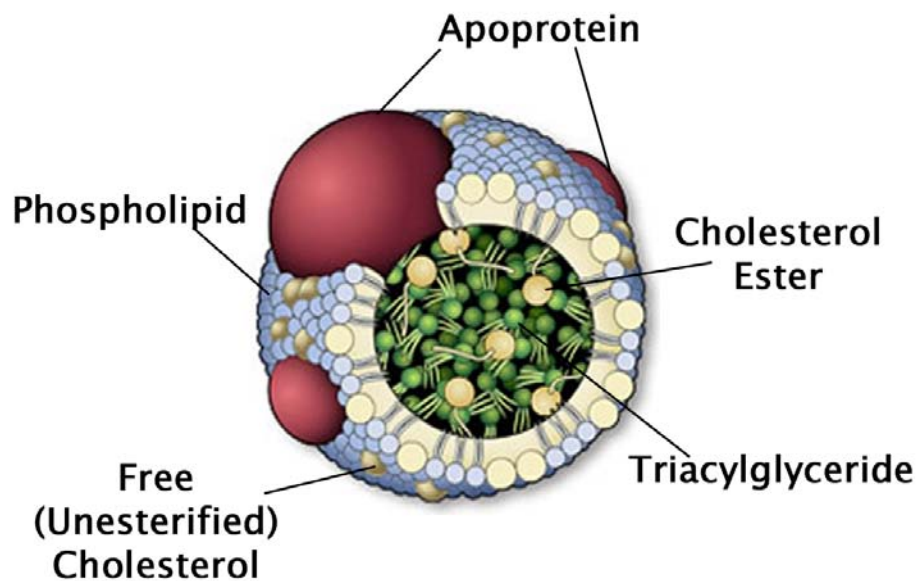


Figure 1. Lipoprotein Structure

HDL and LDL cholesterol levels in the blood are important indicators of many disease states. High blood levels of LDLs are associated with health problems and cardiovascular disease. For this reason, LDL is often referred to as the “bad cholesterol.” LDL particles that accumulate within arteries can form plaques over time, which can increase chances of a stroke, heart attack, or vascular disease. HDL particles are able to remove cholesterol from within arteries and transport it back to the liver for re-utilization or excretion, which is the main reason why the cholesterol carried within HDL particles is sometimes called "good cholesterol." Monitoring circulatory levels of different lipoproteins is critical to the diagnosis of lipid transport disorders such as atherosclerosis.

Cell Biolabs’ HDL-Cholesterol Assay Kit is a simple fluorometric assay that can measure the amounts of HDL cholesterol present in plasma or serum samples within a 96-well microtiter plate format. Each kit provides sufficient reagents to perform up to 96 assays, including blanks, cholesterol standards and unknown samples. Sample cholesterol concentrations are determined by comparison with a known cholesterol standard.

Assay Principle

Cell Biolabs’ HDL-Cholesterol Assay Kit measures the HDL cholesterol levels in serum or plasma samples. The kit provides reagents for separating HDL from other lipoproteins (LDL, VLDL, IDL and Chylomicron) before cholesterol assay. The cholesterol assay is based on the enzyme driven reaction that quantifies both cholesterol esters and free cholesterol. Cholesterol esters are hydrolyzed via cholesterol esterase into cholesterol, which is then oxidized by cholesterol oxidase into the ketone cholest-4-en-3-one and hydrogen peroxide. The hydrogen peroxide is then detected with a highly specific fluorescence probe. Horseradish peroxidase catalyzes the reaction between the probe and hydrogen peroxide, which bind in a 1:1 ratio. Samples are compared to a known concentration of cholesterol standard within the 96-well microtiter plate format. Samples and standards are incubated for 45 minutes and then read with a standard 96-well fluorometric plate reader (Figure 2).

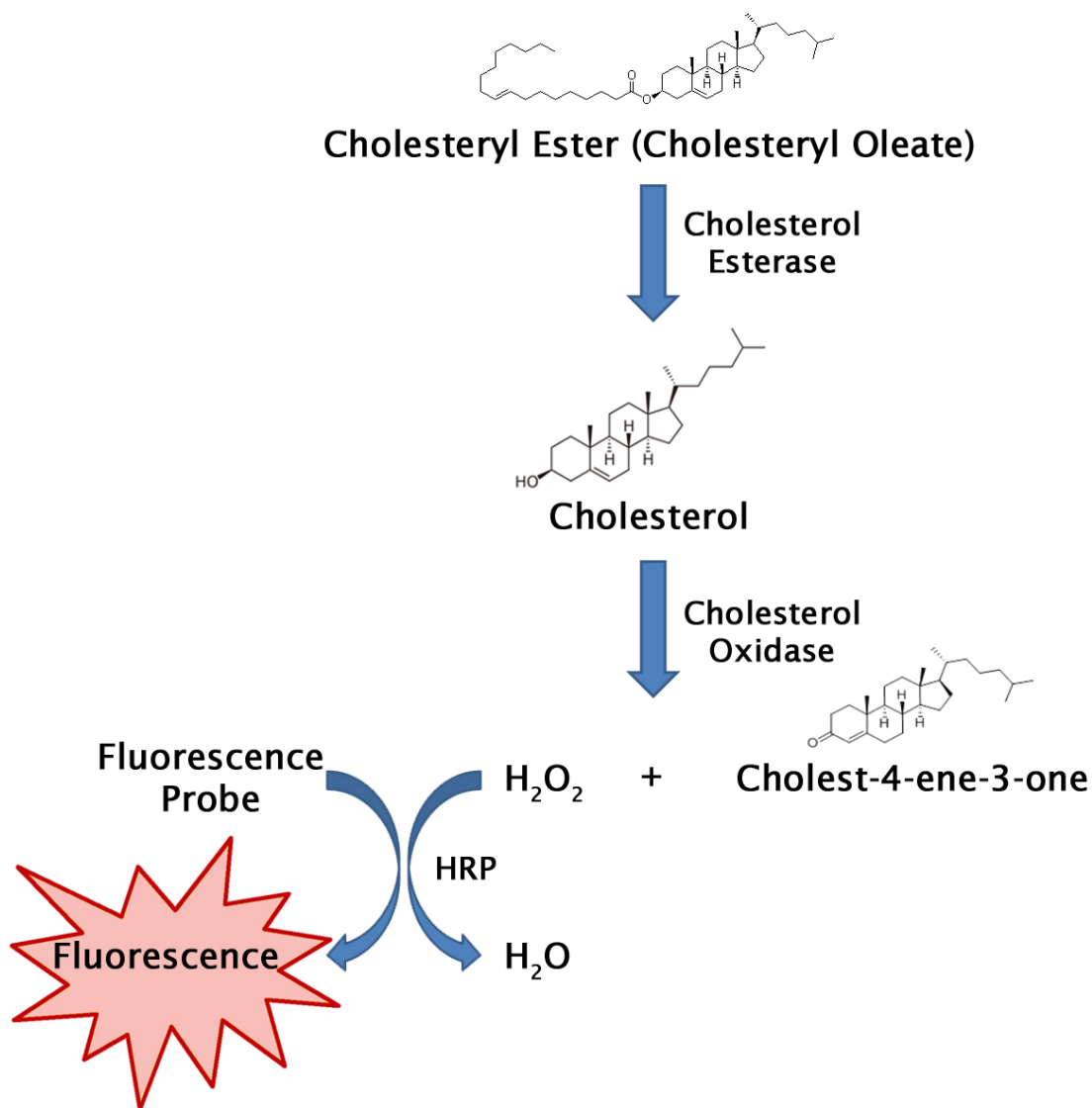


Figure 2. Cholesterol Assay Principle

Related Products

1. STA-241: Human Low Density Lipoprotein (LDL)
2. STA-243: Human High Density Lipoprotein (HDL)
3. STA-359: Human Apo(a) ELISA Kit
4. STA-362: Human ApoAI ELISA Kit
5. STA-363: Human ApoAII ELISA Kit
6. STA-364: Human ApoCI ELISA Kit
7. STA-365: Human ApoCII ELISA Kit

8. STA-366: Human ApoCIII ELISA Kit
9. STA-367: Human ApoE ELISA Kit
10. STA-368: Human ApoB-100 ELISA Kit
11. STA-369: OxiSelect™ Human Oxidized LDL ELISA Kit (MDA-LDL Quantitation)
12. STA-390: Total Cholesterol Assay Kit (Fluorometric)

Kit Components

Box 1 (shipped at room temperature)

1. 96-well Microtiter Plate (Part No. 234501): One 96-well clear bottom black plate.
2. Cholesterol Standard (Part No. 239001): One 50 µL vial of a 10 mM cholesterol solution in ethanol.
3. Assay Diluent (5X) (Part No. 239002): One 100 mL bottle.
4. Fluorescence Probe (Part No. 239005): One 200 µL vial in DMSO.
5. HRP (Part No. 234402): One 100 µL tube of 100 U/mL solution each in glycerol.
6. Precipitation Solution (2X) (Part No. 239401): One 20 mL bottle.

Box 2 (shipped on blue ice packs)

1. Cholesterol Esterase (Part No. 239003): One vial of 10 Units enzyme in powder.
2. Cholesterol Oxidase (Part No. 239004): One 200 µL vial.

Materials Not Supplied

1. Distilled or deionized water
2. Microcentrifuge
3. Microcentrifuge tubes
4. 10 µL to 1000 µL adjustable single channel micropipettes with disposable tips
5. 50 µL to 300 µL adjustable multichannel micropipette with disposable tips
6. Multichannel micropipette reservoir
7. Fluorescence microplate reader capable of reading excitation in the 530-570 nm range and emission in the 590-600 nm range.

Storage

Upon receipt, store the Cholesterol Standard, Fluorescence Probe, HRP, Cholesterol Oxidase, and Cholesterol Esterase at -20°C. The Fluorescence Probe is light sensitive and must be stored accordingly. Avoid multiple freeze/thaw cycles. Store the remaining kit components at 4°C.

Preparation of Reagents

- 1X Assay Diluent: Warm the Assay Diluent (5X) to room temperature prior to using. Dilute the Assay Diluent (5X) with deionized water by diluting the 100 mL Diluent with 400 mL deionized water for 500 mL total. Mix to homogeneity. Store the 1X Assay Diluent at 4°C up to six months.
- Cholesterol Esterase: Reconstitute the powder with 200 μ L of 1X Assay Diluent. Vortex vigorously until dissolved. Prepare aliquots and store at -20°C to avoid multiple freeze thaws of the reconstituted powder.
- Cholesterol Reaction Reagent: Prepare the reagent by diluting the Cholesterol Oxidase 1:50, HRP 1:50, Fluorescence Probe 1:50, and Cholesterol Esterase 1:250 in 1X Assay Diluent. (eg. For 100 assays, combine 100 μ L of Cholesterol Oxidase, 100 μ L of HRP, 100 μ L Fluorescence Probe, and 20 μ L Cholesterol Esterase with 1X Assay Diluent to 5 mL total solution). Mix thoroughly and protect the solution from light. For best results, place the Cholesterol Reaction Reagent on ice and use within 30 minutes of preparation. Do not store the Cholesterol Reaction Reagent solution.

Note: The Fluorescence Probe is light sensitive and must be stored accordingly.

Preparation of Samples

Samples should be used immediately or stored at -80°C prior to performing the assay. Optimal experimental conditions for samples must be determined by the investigator. The following recommendations are only guidelines and may be altered to optimize or complement the user's experimental design. A set of serial dilutions is recommended for samples to achieve optimal assay results and minimize possible interfering compounds. Run proper controls as necessary. Always run a standard curve with samples.

- Serum: Collect blood in a tube with no anticoagulant. Allow the blood to clot at room temperature for 30 minutes. Centrifuge at 2500 x g for 20 minutes. Remove the serum layer and store on ice. Avoid disturbing the white buffy layer. Aliquot samples for testing and store at -80°C. Perform dilutions in 1X Assay Diluent. Cholesterol levels in serum average about 3% higher in value than in the corresponding plasma pair (Ref. 2).
- Plasma: Avoid hemolyzed and lipemic blood samples. Collect blood with heparin or citrate and centrifuge at 2000 x g and 4°C for 10 minutes. Remove the plasma layer and store on ice. Avoid disturbing the white buffy layer. Aliquot samples for testing and store at -80°C. Perform dilutions in 1X Assay Diluent.

Preparation of HDL Fraction

1. Add 200 μ L of sample (serum or plasma) to a microcentrifuge tube. Add 200 μ L of the 2X Precipitation Reagent and mix well by vortexing. Allow mixture to incubate 5-10 minutes at room temperature (precipitation will occur).
2. Centrifuge the mixture at 2000 x g (~5000 rpm) for 20 minutes (pellet should be visible). Slowly and carefully transfer the supernatant (HDL fraction) into a new tube, discard the pellet (other lipoproteins including LDL, VLDL and Chylomicron).
3. Further dilute the HDL fraction samples 1:25 (1:50 final dilution) to 1:100 (1:200 final dilution) in 1X Assay Diluent before running the assay. Assay immediately and do not store solutions.

Preparation of Cholesterol Standard Curve

Prepare fresh cholesterol standards by diluting in 1X Assay Diluent. First, dilute the stock Cholesterol Standard 10 mM solution 1:50 in 1X Assay Diluent for a 200 μ M solution. (eg. add 20 μ L of the stock 10 mM standard to 980 μ L of 1X Assay Diluent). Vortex thoroughly. Use this 200 μ M solution to prepare a series of the remaining cholesterol standards according to **Table 1** below.

Tubes	200 μM Cholesterol Standard (μL)	1X Assay Diluent (μL)	Resulting Cholesterol Concentration (μM)
1	60	940	12
2	50	950	10
3	40	960	8
4	30	970	6
5	20	980	4
6	10	990	2
7	5	995	1
8	0	1000	0

Table 1. Preparation of Cholesterol Standards.

Note: Do not store diluted cholesterol standard solutions.

Assay Protocol

Each cholesterol standard and sample should be assayed in duplicate or triplicate. A freshly prepared standard curve should be used each time the assay is performed.

1. Add 50 μ L of the diluted cholesterol standards or the diluted HDL fraction samples to the 96-well microtiter plate.
2. Add 50 μ L of the prepared Cholesterol Reaction Reagent to each well and mix the well contents thoroughly.
3. Cover the plate wells to protect the reaction from light. Incubate the plate for 45 minutes at 37°C.
4. IMMEDIATELY read the plate with a fluorescence microplate reader equipped for excitation in the 530-570 nm range and for emission in the 590-600 nm range.
5. Calculate the concentration of cholesterol within samples by comparing the sample RFU to the cholesterol standard curve.

Example of Results

The following figures demonstrate typical HDL-Cholesterol Assay results. One should use the data below for reference only. This data should not be used to interpret or calculate actual sample results.

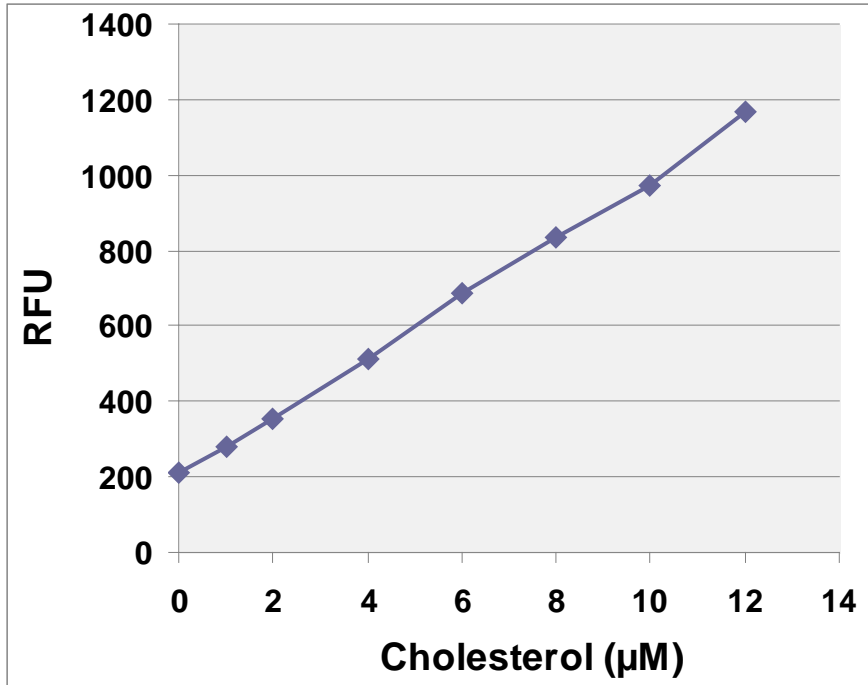


Figure 3: Cholesterol standard curve.

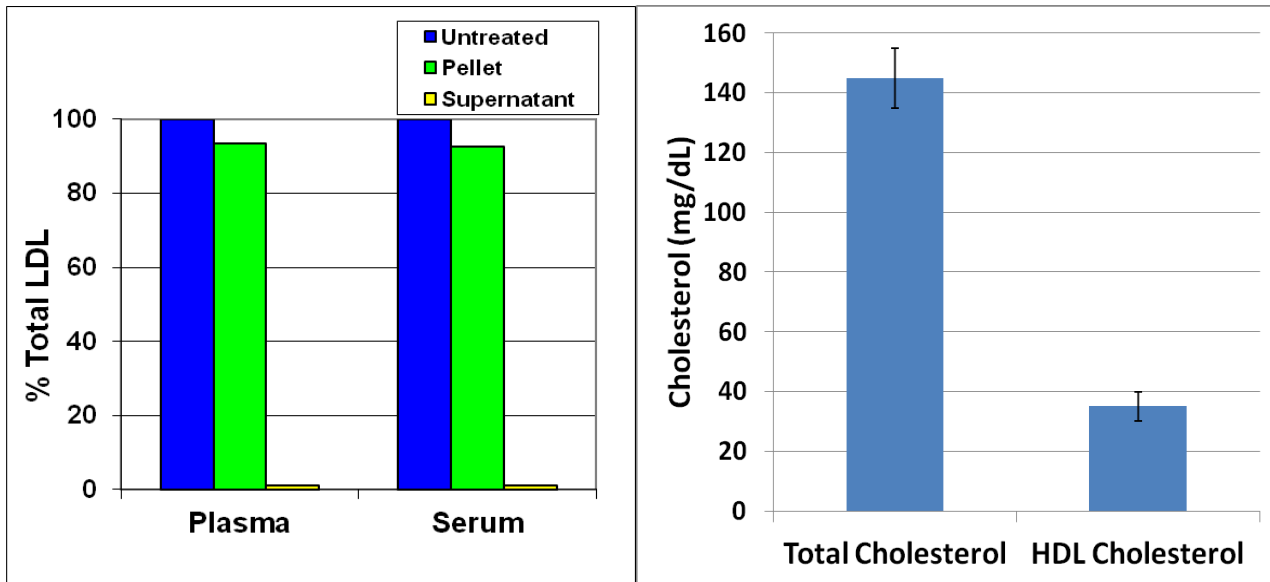


Figure 4: HDL cholesterol values in human serum. Left: LDL/VLDL removal after precipitation. Serum and plasma samples were treated with Precipitation Solution according to the Sample Preparation protocol. LDL/VLDL removal was determined by Human ApoB ELISA (STA-368). **Right:** HDL-Cholesterol Assays on human serum before and after precipitation.

Calculation of Results

1. Calculate the average fluorescence values for every standard, control, and sample. Subtract the average zero standard value from itself and all standard and sample values. This is the corrected fluorescence.
2. Plot the corrected fluorescence for the standards against the final concentration of the cholesterol standards from Table 1 to determine the best curve. See Figure 3 for an example standard curve.
3. Determine the cholesterol concentration of the HDL samples with the equation obtained from the linear regression analysis of the standard curve. Substitute the corrected fluorescence values for each sample. Remember to account for all dilution factors.

$$\text{HDL Cholesterol } (\mu\text{M}) = \left[\frac{\text{Sample corrected fluorescence}}{\text{Slope}} \right] \times \text{Sample dilution}$$

Note: For the conversion of results from μM to mg/dl , divide the cholesterol concentration (μM) by 25.9.

References

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4. Ledwozyw, A., et al. (1986) *Clin. Chim. Acta.* **155**: 275-284.
5. Lee, S.M. et al. (2008) *Lipids* **43**: 419-429.

Warranty

These products are warranted to perform as described in their labeling and in Cell Biolabs literature when used in accordance with their instructions. THERE ARE NO WARRANTIES THAT EXTEND BEYOND THIS EXPRESSED WARRANTY AND CELL BIOLABS DISCLAIMS ANY IMPLIED WARRANTY OF MERCHANTABILITY OR WARRANTY OF FITNESS FOR PARTICULAR PURPOSE. CELL BIOLABS's sole obligation and purchaser's exclusive remedy for breach of this warranty shall be, at the option of CELL BIOLABS, to repair or replace the products. In no event shall CELL BIOLABS be liable for any proximate, incidental or consequential damages in connection with the products.

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