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**Product Manual**

# **ROCK Activity Immunoblot Kit**

**Catalog Number**

**STA-415**

**20 assays**

**FOR RESEARCH USE ONLY**  
**Not for use in diagnostic procedures**

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## **Introduction**

Members of the Rho family are essential regulatory components of the signaling pathway that direct cell motility, adhesion, and cytokinesis through reorganization of actin cytoskeleton. Rho is activated by extracellular signals such as lysophosphatidic acid (LPA). The actions of Rho are mediated by downstream Rho effectors. One of these effectors is Rho-associated kinase (ROCK). Two ROCK isoforms have been identified: ROCK-I (also known as ROK $\beta$ ) and ROCK-II (also known as Rho Kinase and ROK $\alpha$ ). ROCK mediates Rho signaling and reorganizes actin cytoskeleton through phosphorylation of several substrates that contribute to the assembly of actin filaments and contractility. For example, ROCK inactivates myosin phosphatase through the specific phosphorylation of myosin phosphatase target subunit 1 (MYPT1) at Thr<sup>696</sup>, which results in an increase in the phosphorylated content of the 20-kDa myosin light chain (MLC20).

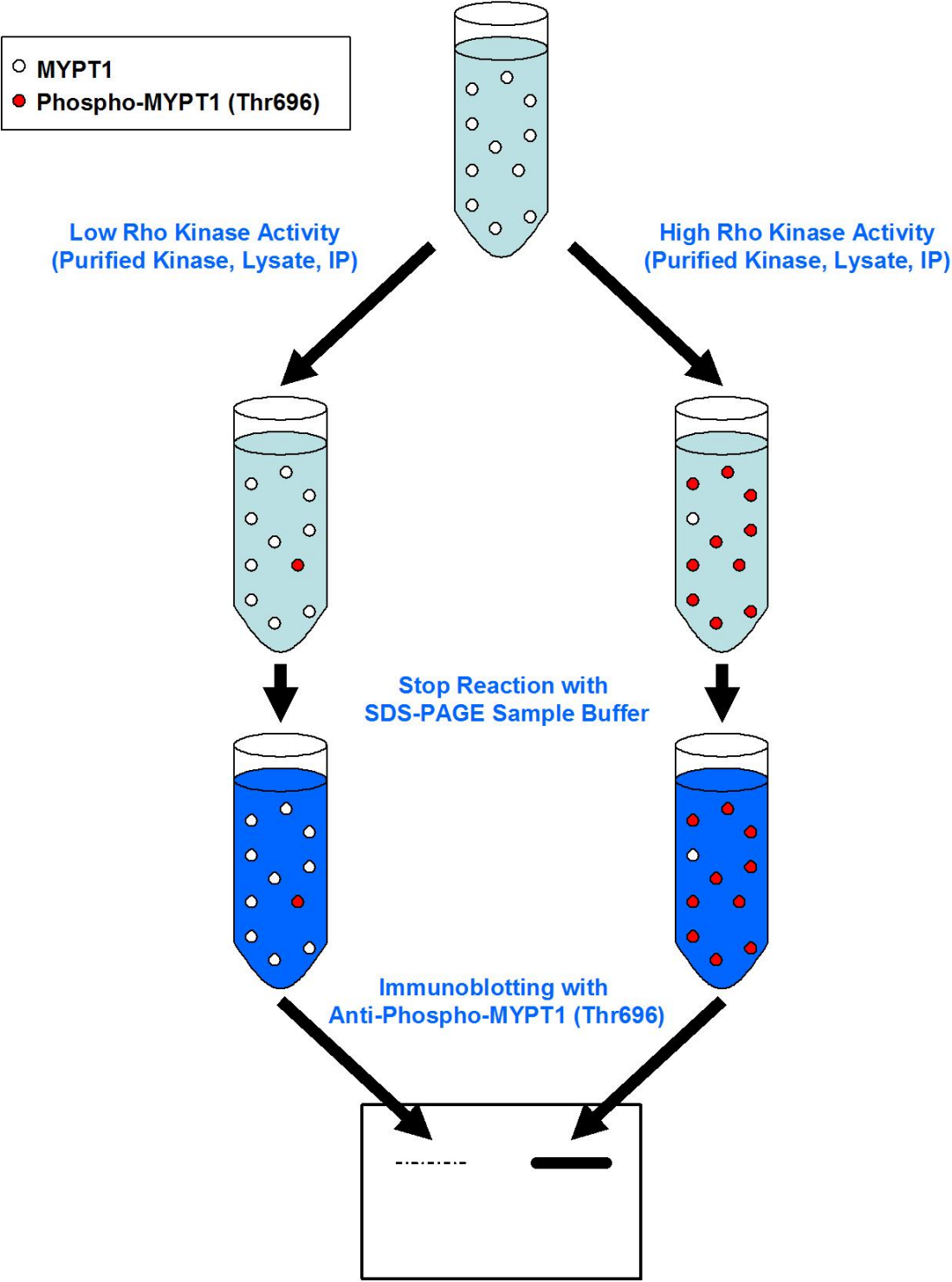
Cell Biolabs' ROCK Activity Immunoblot Kit utilizes recombinant MYPT1 as ROCK substrate. After incubating the substrate with ROCK samples (such as purified kinase, cell lysate or immunoprecipitate), the phosphorylated MYPT1 is detected by western blot analysis using an anti-phospho-MYPT1 (Thr<sup>696</sup>) (Figure 1).

Cell Biolabs' ROCK Activity Immunoblot Kit provides a simple and fast tool to monitor ROCK activity using its physiological substrate. The kit also includes active ROCK-II as a positive control. Each kit provides sufficient quantities to perform 20 assays.

## **Related Products**

1. STA-416: 96-well ROCK Activity Assay Kit
2. STA-400: Ras Activation Assay Kit
3. STA-402: Cdc42 Activation Assay Kit
4. STA-403: Rho Activation Assay Kit
5. STA-404: Rac/Cdc42 Activation Assay Combo Kit
6. STA-405: Rho/Rac/Cdc42 Activation Assay Combo Kit
7. STA-410: PAK1 PBD Agarose Beads
8. STA-411: Raf1 PBD Agarose Beads
9. STA-412: Rhotekin PBD Agarose Beads
10. STA-452: GFP-RhoA Expression Vector Set
11. STA-456: RhoA Expression Vector Set
12. STA-460: Exoenzyme C3 (Rho Inhibitor) Expression Vector

**Assay Principle**



## **Kit Components**

1. **ROCK Substrate** (Part No 241501): One 40  $\mu$ L vial containing 0.25 mg/mL recombinant MYPT1
2. **10X Kinase Buffer** (Part No. 241502): Three 1.0 mL vials of 250 mM Tris, pH 7.5, 100 mM  $MgCl_2$ , 50 mM Glycerol-2-Phosphate, 1 mM  $Na_3VO_4$
3. **ATP Solution** (Part No. 241503): One 400  $\mu$ L vial of 10 mM ATP
4. **Anti-phospho-MYPT1 (Thr<sup>696</sup>)** (Part No. 241504): One 50  $\mu$ L vial
5. **Secondary Antibody, HRP-conjugate** (Part No. 230805): One 100  $\mu$ L vial
6. **Active ROCK-II** (Part No. 241505): One 20  $\mu$ L vial containing 10 ng active ROCK-II in 25 mM Tris, pH 7.5, 10 mM  $MgCl_2$ , 5 mM Glycerol-2-Phosphate, 0.1 mM  $Na_3VO_4$ , 10% Glycerol, 0.1% BSA

## **Materials Not Supplied**

1. ROCK sample (purified kinase, cell lysate or immunoprecipitate)
2. DTT
3. 30°C incubator or water bath
4. 4X SDS-PAGE sample buffer
5. Electrophoresis and immunoblotting systems
6. Immunoblotting wash buffer such as TBST (10 mM Tris-HCl, pH 7.4, 0.15 M NaCl, 0.05% Tween-20)
7. Immunoblotting blocking buffer (TBST containing 5% Non-fat Dry Milk)
8. PVDF or nitrocellulose membrane
9. ECL Detection Reagents

## **Storage**

Store active ROCK-II at -80°C and all other kit components at -20°C. Avoid multiple freeze/thaw cycles.

## **Preparation of Reagents**

- 1X Kinase Buffer: Dilute to 10X Kinase Buffer to 1X in deionized water. 1X Kinase Buffer may be stored at 4°C for short term (1-2 weeks). Just prior to usage, add DTT to a final concentration of 1 mM.
- 1X Kinase/ATP/Substrate Solution: For each kinase assay, freshly prepare 50  $\mu$ L of 1X Kinase/ATP/Substrate Solution by adding 1  $\mu$ L of 10 mM ATP solution, 2  $\mu$ L of ROCK substrate to 47  $\mu$ L of 1X Kinase Buffer containing DTT.

## **Assay Protocol**

### **I. Kinase Reaction**

- 1a. For Immunoprecipitations with anti-ROCK antibody: ROCK is first immunoprecipitated from cell or tissue lysate with anti-ROCK antibody and Protein A/G bead slurry. Immediately before kinase assay, wash bead slurry once with 1X Kinase Buffer, remove all supernatant, assay immediately by adding 50  $\mu$ L of 1X Kinase/ATP/Substrate Solution directly to the beads and mixing well.
- 1b. Purified Kinase or Cell Lysate: Purified kinase or cell lysate sample can be used directly in the kinase assay or further diluted with 1X Kinase Buffer. Add 25  $\mu$ L of ROCK sample to a microcentrifuge tube, and initiate kinase reaction by adding 50  $\mu$ L of 1X Kinase/ATP/Substrate Solution to the ROCK sample.
2. (optional) Add 2  $\mu$ L of the provided active ROCK-II and 23  $\mu$ L of 1X Kinase Buffer to a microcentrifuge tube, initiate kinase reaction by adding 50  $\mu$ L of 1X Kinase/ATP/Substrate Solution.
3. Incubate the tubes at 30°C for 30-60 minutes with gentle agitation.
4. Stop kinase reaction by adding 25  $\mu$ L of 4X reducing SDS-PAGE sample buffer.
5. Boil each sample for 5 minutes.
6. Centrifuge each sample for 10 seconds at 12,000 x g.

### **II. Electrophoresis and Transfer**

1. Load 20  $\mu$ L of supernatant to a polyacrylamide gel. Also, it's recommended to include a pre-stained MW standard (as an indicator of a successful transfer in step 3).
2. Perform SDS-PAGE as per the manufacturer's instructions.
3. Transfer the gel proteins to a PVDF or nitrocellulose membrane as per the manufacturer's instructions.

### **III. Immunoblotting and Detection (all steps are at room temperature, with agitation)**

1. Following the electroblotting step, immerse the PVDF membrane in 100% Methanol for 15 seconds, and then allow it to dry at room temperature for 5 minutes.

*Note: If Nitrocellulose is used instead of PVDF, this step should be skipped.*

2. Block the membrane with 5% non-fat dry milk in TBST for 1 hr at room temperature with constant agitation.
3. Incubate the membrane with the anti-phospho-MYPT1 (Thr<sup>696</sup>) antibody, freshly diluted 1:1000 in 5% non-fat dry milk/TBST, for 2 hr at room temperature with constant agitation.

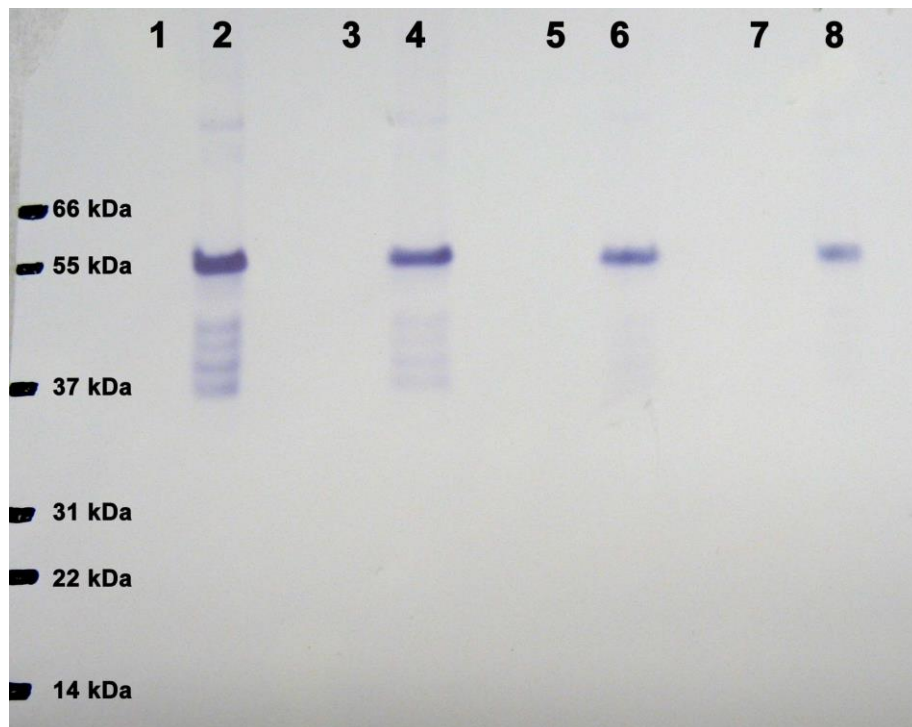
*Note: To conserve antibody, incubations should be performed in a plastic bag.*

4. Wash the blotted membrane three times with TBST, 5 minutes each time.
5. Incubate the membrane with the secondary antibody, HRP-conjugated, freshly diluted in 1:1000 in 5% non-fat dry milk/TBST, for 1 hr at room temperature with constant agitation.

6. Wash the blotted membrane three times with TBST, 5 minutes each time.
7. Use the detection method of your choice. We recommend enhanced chemiluminescence reagents from Pierce.

### **Example of Results**

The following figure demonstrates typical results seen with Cell Biolabs ROCK Activity Immunoblot Kit. One should use the data below for reference only.



**Figure 1: ROCK-II Activity Immunoblot Assay.** 25  $\mu$ L of 1X Kinase Buffer containing 10 ng of active ROCK-II was incubated with 50  $\mu$ L of 1X Kinase Buffer containing 0.2 mM ATP and 500 ng of recombinant MYPT1 for 30 minutes at 30°C. Kinase reaction was stopped by adding 25  $\mu$ L of 4X SDS-PAGE Sample Buffer. Lane 1, 3, 5, 7: Without kinase (negative control); Lane 2, 4, 6, 8: with kinase. 200 ng (Lane 1 and 2), 100 ng (Lane 3 and 4), 50 ng (Lane 5 and 6) or 25 ng (Lane 7 and 8) of recombinant MYPT1 substrate were loaded onto SDS-PAGE. Phosphorylation of MYPT1 substrate was detected by anti-phospho-MYPT1 (Thr<sup>696</sup>) antibody as described in Assay Protocol.

### **References**

1. Etienne-Manneville, S., and Hall, A. (2002) *Nature* **420**, 629-635.
2. Riento, K., and Ridley, A. J. (2003) *Nat. Rev. Mol. Cell. Biol.* **4**, 446-456.
3. Leung, T., Manser, E., Tan, L., and Lim, L. (1995) *J. Biol. Chem.* **270**, 29051-29054.
4. Matsui, T., Amano, M., Yamamoto, T., Chihara, K., Nakafuku, M., Ito, M., Nakano, T., Okawa, K., Iwamatsu, A., and Kaibuchi, K. (1996) *EMBO J.* **15**, 2208-2216.

5. Totsukawa, G, *et al.*, *J. Cell Biol.* (1999) **144**, 735-744.

## **Recent Product Citations**

1. Rothschild, P.R., *et al.* (2017). ROCK-1 mediates diabetes-induced retinal pigment epithelial and endothelial cell blebbing: Contribution to diabetic retinopathy. *Sci Rep.* **7**(1):8834. doi: 10.1038/s41598-017-07329-y.
2. Watts, B. A. *et al.* (2016). High mobility group box 1 inhibits HCO<sub>3</sub>-absorption in medullary thick ascending limb through RAGE-Rho-ROCK-mediated inhibition of basolateral Na<sup>+</sup>/H<sup>+</sup> exchange. *Am J Physiol Renal Physiol.* doi:10.1152/ajprenal.00185.2016.
3. Seo, J. H. *et al.* (2016). Dynamic polyrotaxane-coated surface for effective differentiation of mouse induced pluripotent stem cells into cardiomyocytes. *RSC Adv.* doi:10.1039/C6RA03967G.
4. Tocci, P. *et al.* (2016). Endothelin-1/endothelin A receptor axis activates RhoA GTPase in epithelial ovarian cancer. *Life Sci.* doi:10.1016/j.lfs.2016.01.008.
5. Su, C. C. *et al.* (2015). Phenotypes of trypsin-and collagenase-prepared bovine corneal endothelial cells in the presence of a selective Rho kinase inhibitor, Y-27632. *Mol Vis.* **21**:633-643.
6. Gross, P. *et al.* (2015). Para-cresyl sulfate acutely impairs vascular reactivity and induces vascular remodeling. *J Cell Physiol.* doi: 10.1002/jcp.25018.
7. Sailland, J. *et al.* (2014). Estrogen-related receptor  $\alpha$  decreases RHOA stability to induce orientated cell migration. *Proc Natl Acad Sci U S A.* **111**:15108-15113.
8. Georgess, D. *et al.* (2014). Comparative transcriptomics reveals RhoE as a novel regulator of actin dynamics in bone-resorbing osteoclasts. *Mol. Biol.* **25**:380-396.
9. Chandrasekharan, U. *et al.* (2013). Release of nonmuscle myosin II from the cytosolic domain of tumor necrosis factor receptor 2 is required for target gene expression. *Sci. Signal.* **6**:ra60.
10. Gupta, N. *et al.* (2012). Soluble epoxide hydrolase: sex differences and role in endothelial cell survival. *Arterioscler Thromb Vasc Biol.* **32**:1936-1942.
11. Forero, D.G. *et al.* (2012). Endogenous Rho-kinase signaling maintains synaptic strength by stabilizing the size of the readily releasable pool of synaptic vesicles. *J. Biol. Chem.* **287**:1600-1608.
12. Wang, J.N. *et al.* (2011). Response gene to complement 32 promotes vascular lesion formation through stimulation of smooth muscle cell proliferation and migration. *Arterioscler Thromb Vasc Biol.* **31**:e19-e26.
13. Li, Z. *et al.* (2009). TrkB1 induces liver metastasis of pancreatic cancer cells by sequestering Rho GDP dissociation inhibitor and promoting RhoA activation. *Cancer Res.* **69**:7851-7859.
14. Xiao, L. *et al.* (2009). ROCK mediates phorbol ester-induced apoptosis in prostate cancer cells via p21-Cip1 upregulation and JNK. *J. Biol. Chem.* **284**:29365-29375.

## **Warranty**

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