

pAAV-MCS Expression Vector

CATALOG NUMBER: VPK-410

STORAGE: -20°C

QUANTITY AND CONCENTRATION: 10 µg at 0.25 µg/µL in TE

Background

Adeno-associated viruses (AAVs) are derived from defective parvoviruses, which depend on essential helper functions provided by other viruses, such as adenovirus and herpes virus, for efficient viral replication and propagation. AAV has no etiologic association with any known diseases and has been successfully used to establish efficient and long-term gene expression *in vivo* in a variety of tissues without significant cellular immune responses or toxicity.

AAV has a single-stranded DNA genome which consists of approximately 4.7 kb. All characterized AAV serotypes share three key features, including two copies of AAV terminal repeats (ITRs), one *rep* region and one *cap* region. The ITRs are capable of forming T-shape secondary structure and are the only *cis* elements that are required for AAV replication, packaging, integration, and rescue. The *rep* region encodes four overlapping proteins designated as Rep78, Rep68, Rep52, and Rep40, according to the apparent molecular mass of the protein. In addition to their well-defined roles in AAV replication, Rep proteins also regulate AAV packaging and site-specific integration. The *cap* region encodes three structural proteins, VP1, VP2, and VP3. These three proteins share the same reading frame (see Figure 1).

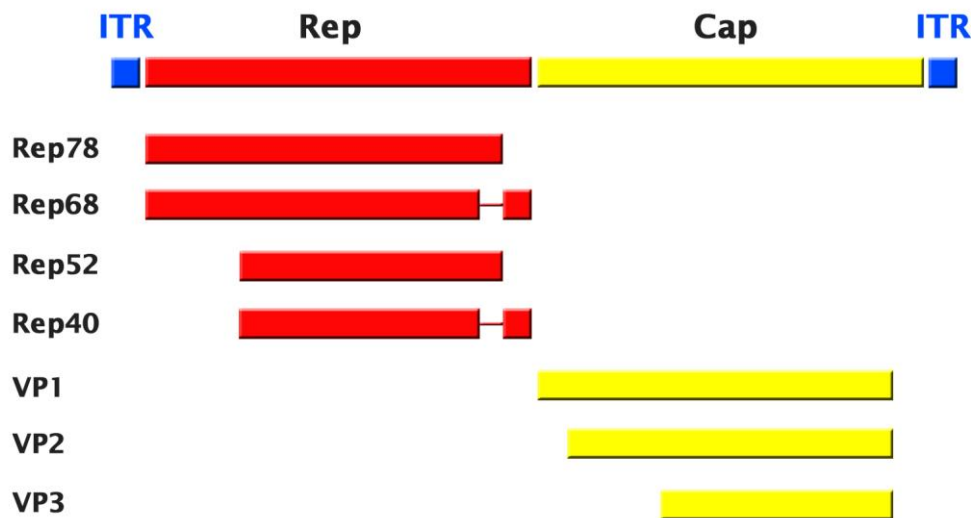


Figure 1. Schematic Map of AAV Genome. Rep: involved in genome replication; VP1/2/3: capsid proteins.

Cell Biolabs' AAV Helper-Free System allows the production of infectious recombinant human adeno-associated virus (rAAV) virions without the use of a helper virus (Figure 2). In the AAV Helper-Free System, most of the adenovirus gene products required for the production of infective AAV particles are supplied on the plasmid pHelper (i.e. E2A, E4, and VA RNA genes) that is co-transfected into cells with human AAV vector DNA. The remaining adenoviral gene product is supplied by the 293 host

cells, which stably express the adenovirus E1 gene. By eliminating the requirement for live helper virus the AAV Helper-Free System provides a safer and more convenient gene delivery system. In the AAV Helper-Free System, the *rep* and *cap* genes have been removed from the viral vector that contains AAV-2 ITRs and are supplied in *trans* on the plasmid pAAV-RC. The removal of the AAV *rep* and *cap* genes allows for insertion of a gene of interest in the viral genome.

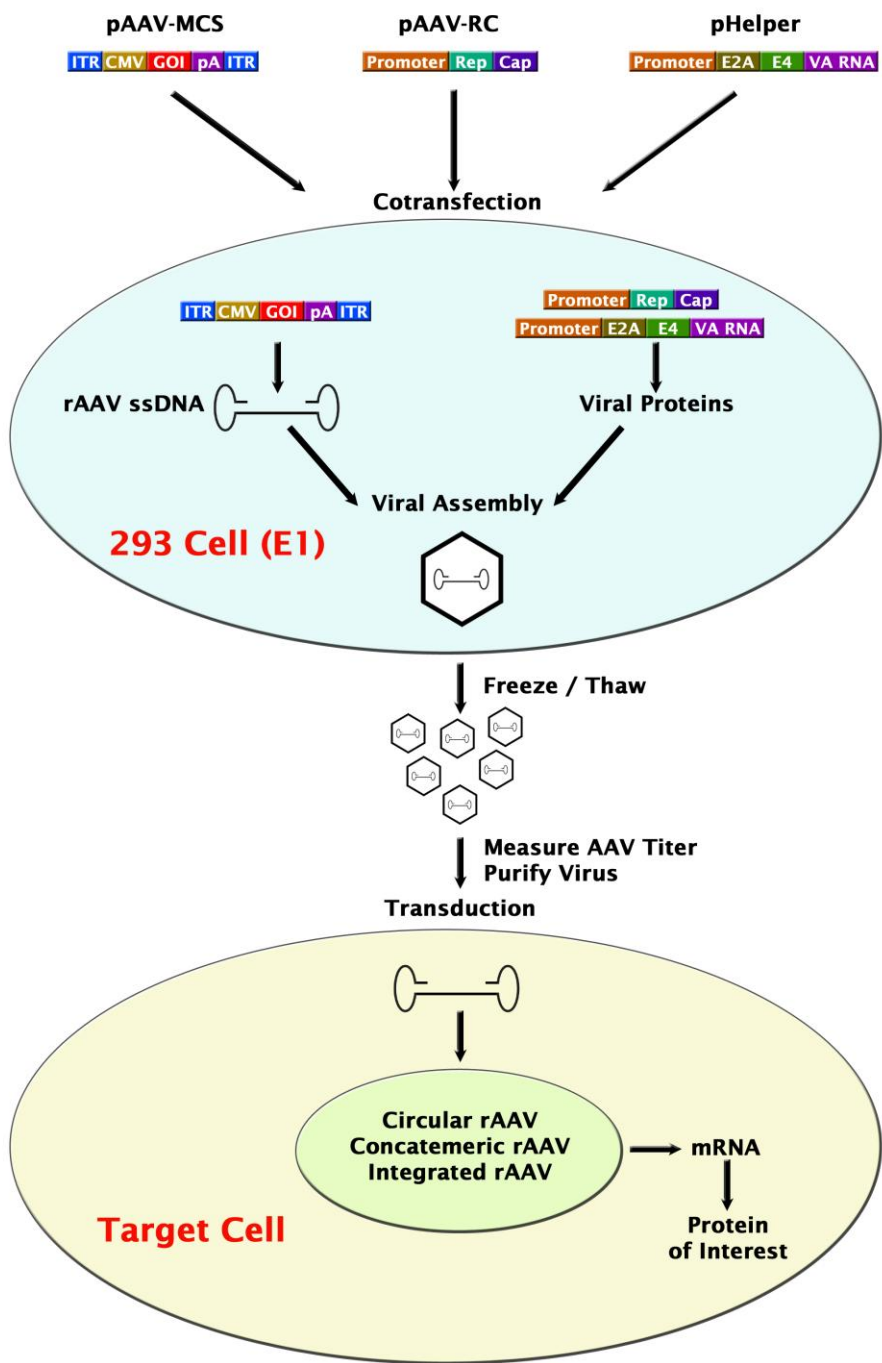


Figure 2. AAV Helper-Free system.

Catalog #	Product Name	Capacity (kb)
VPK-410	pAAV-MCS	3
VPK-411	pAAV-MCS	3.9
VPK-418	pAAV-IRES-GFP	1.7

Table 1. Packaging capacity of AAV shuttle vectors.

Recombinant adeno-associated viruses are important tools for gene delivery and expression. AAV has not been reported to cause any diseases. Together with its replication defective nature, AAV has good safety profile to be used in gene transfer in vivo, and as potential gene therapy vehicles. Recombinant AAV is capable of infecting a broad range of cell types including non-dividing cells and remaining as concatemers for long-term expression. Compared with other viral vectors such as adenovirus, AAV elicits very mild immune response in animal models.

Related Products

1. VPK-402: AAV Helper Free Packaging System
2. AAV-100: 293AAV Cell Line
3. VPK-140: ViraBind™ AAV Purification Kit
4. VPK-145: QuickTiter™ AAV Quantitation Kit
5. AAV-200: ViraDuctin™ AAV Transduction Kit

Safety Considerations

Remember that you will be working with samples containing infectious virus. Follow the recommended NIH guidelines for all materials containing BSL-2 organisms. The AAV Helper-Free system is designed to minimize the chance of generating wild type AAV, but precautions should still be taken to avoid direct contact with viral supernatants.

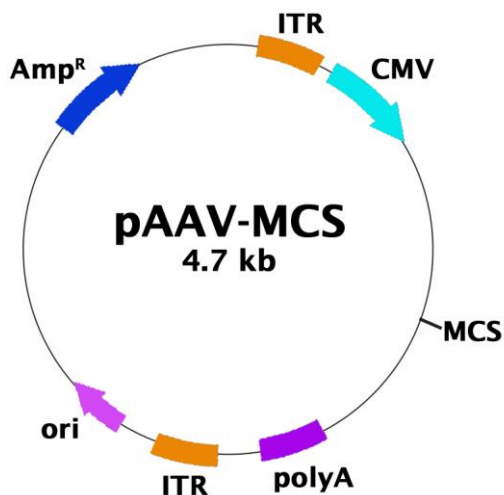


Figure 3: pAAV-MCS Expression Vector (see Appendix for more detail).

Vector Features:

- 1 ~ 130: Left ITR
- 150 ~ 812: CMV Promoter
- 820 ~ 1312: human β -globin intron
- 1319 ~ 1394: MCS
- 1395 ~ 1873: PolyA
- 1913 ~ 2053: Right ITR
- 2970 ~ 3827: Ampicillin Resistance

MCS:

- Enzyme Sites: 5'- ClaI, EcoRI, BamHI, Sall, HindIII, XhoI (2), BglII -3'

- MCS Sequence:

AACATCGATTGAATTCCCCGGGGATCCTCTAGAGTCGACCTGCAGAAGCTTGCCTCGAGCAGCGCTGCTCGAGAGATCTACG

rAAV Production

1. One day before transfection, plate sufficient 293 cells or 293AAV cells (Cat. # AAV-100) to achieve 70-80% confluence on the day of transfection.
2. Cotransfect cells with pAAV Expression vector, pAAV-RC and pHelper.
Notes:
 - *We recommend the ratio of vectors at 1:1:1 (pAAV Expression Vector:pAAV-RC:pHelper).*
 - *Calcium Phosphate transfection method is preferred for AAV production. For lipid-based transfection reagents, we only suggest FuGENE® 6 (Roche Applied Science) or Lipofectamine™ LTX (Invitrogen).*
3. 48-72 hours after transfection, add 0.5 M EDTA to a final of 10 mM to the plate and incubate for 3 min at room temperature. Gently shake the culture plate several times and harvest all media, including cells, in a sterile tube.
Notes:
 - *As viral production proceeds, some of the cells will round up and detach from the plate, and can be seen as floating in the medium.*
 - *Viruses are present in both intact cells and the growth medium. For more concentrated virus stock, we only recommend proceeding with cell pellet.*
4. Centrifuge the cell suspension at 1000 RPM for 5 min. Remove the supernatant and resuspend the cell pellet in desired amount of DMEM or sterile PBS.
5. Freeze and thaw the cell suspension four times by placing it alternately in a dry ice/ethanol bath and a water bath of 37°C. Remove cell debris by centrifugation at 10,000 g for 10 min and collect the supernatant as AAV crude lysate.
6. AAV crude lysate can be used directly or purified/concentrated if needed. For long term storage, store supernatant at -80°C in aliquots.

Post-Packaging Considerations

The quality of rAAV vector preparations is a key element in obtaining reliable and reproducible data. Purification of rAAV from crude cell lysate is usually required before transduction of your target cells. rAAV is usually quantified by genome copy (GC) number. These genome-containing particles are functional vectors that infect target cells. Besides these "full" AAV, empty viral particles are also produced. Measurement of GC rather than total particle number is thus more relevant.

1. **Concentration and purification of your rAAV:** Recombinant AAV vector can be purified by CsCl gradient ultracentrifugation, iodixanol discontinuous gradient ultracentrifugation, and high-performance liquid chromatography (HPLC). For AAV-2, we recommend using Cell Biolabs' ViraBind™ AAV Purification Kit (Catalog # VPK-140).
2. **Measure titer of your rAAV:**
 - a. **Genome Copy (GC) Number:** This is an important step to ensure consistent viral transduction into your host cell. However, QPCR or dot blot of viral DNA can take as much as 1-4 days to complete. An ELISA method has been developed by using antibody that only reacts with AAV intact particles; however, this method measures all AAV particles including the ones lacking

genomic DNA. Cell Biolabs' QuickTiter™ AAV Quantitation Kit (Catalog # VPK-145) does not involve cell infection; instead it specifically measures the viral nucleic acid content of purified viruses or unpurified viral supernatant sample. The entire procedure takes about 4 hours for unpurified supernatant or about 30 minutes for purified AAV.

- b. Infectious Titer: For AAV vector containing reporter, the rAAV infectious titer can be determined using either green fluorescent protein (GFP) or LacZ as the reporter gene. For rAAV-LacZ, each blue cell after X-Gal staining represents one infectious unit (IU). For rAAV-GFP, each green cell under fluorescence microscopy represents one IU.

- 3. **Use transduction reagents to increase infection efficiency:** The AAV transduction process includes viral binding and entry, intracellular trafficking, nuclear transport, and viral second strand DNA synthesis. The viral second strand DNA synthesis has been shown to be the rate limiting step, which leads to inefficient transduction by AAV vectors. Cell Biolabs' ViraDuctin™ AAV Transduction Kit (Catalog # AAV-200) is designed to increase transduction efficiencies by AAV on both dividing and non-dividing cells.

Example of Results

The following figure demonstrates typical results seen with Cell Biolabs' AAV Helper-Free System. One should use the data below for reference only.

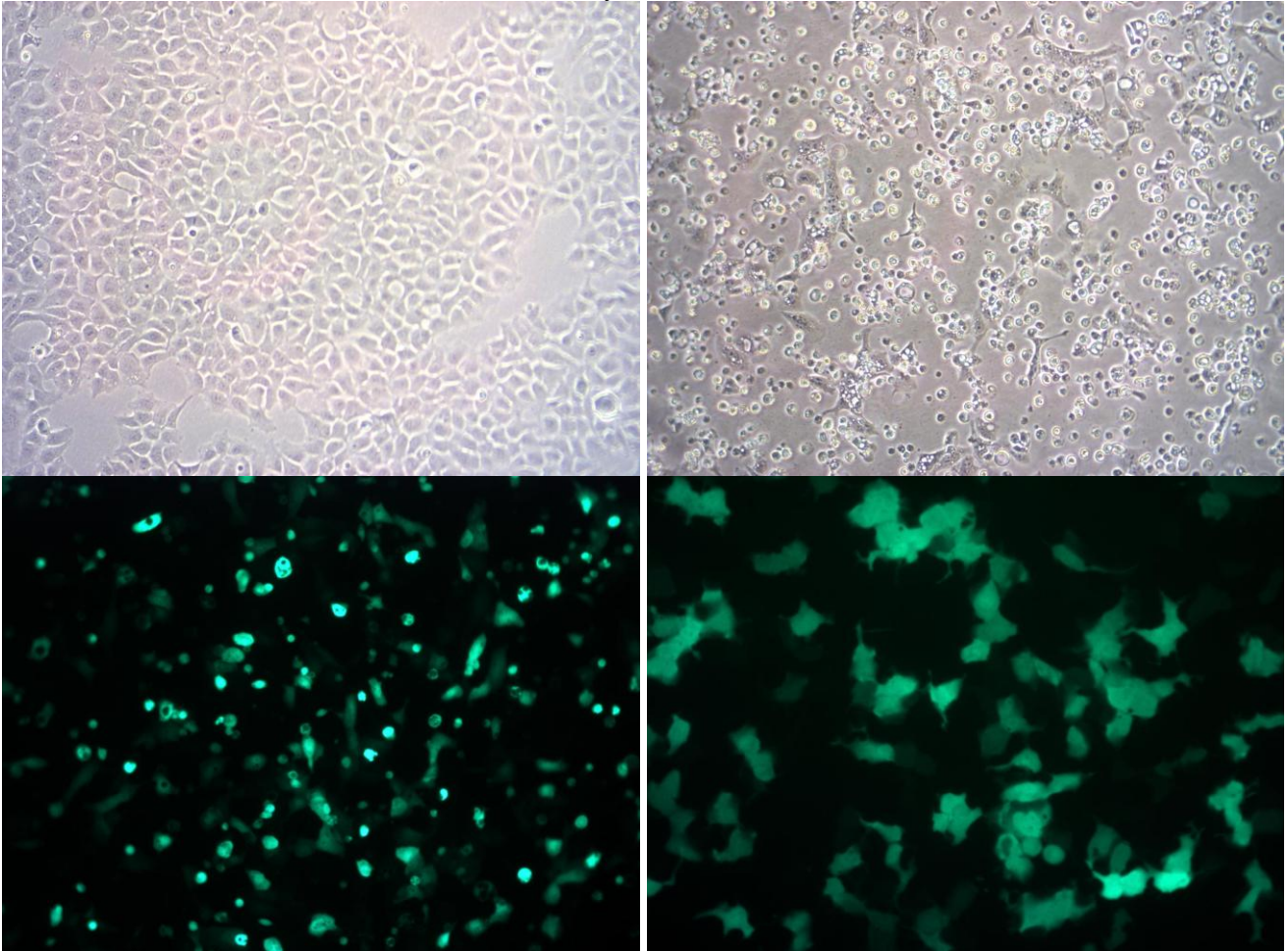


Figure 4: AAV2-GFP Production and Transduction: AAV2-GFP is produced by cotransfecting 293AAV cells (Cat.# AAV-100) with pAAV-GFP (Cat.# AAV-400), pAAV-RC2 and pHelper. 293AD Cells (Cat.# AD-100) were infected with AAV2-GFP viral supernatant for 48 hrs. **Top left:** 293AAV cells before transfection (10X); **Top right:** 293AAV cells 48 hrs after transfection (10X); **Bottom left:** GFP Expression in 293AAV cells 48 hrs after transfection (10X). **Bottom right:** GFP Expression in 293AD cells 48 hrs after transduction (20X).

References

1. Auricchio, A., Hildinger, M., O'Connor, E., Gao, G. P. and Wilson, J. M. (2001) *Hum Gene Ther* **12**:71–6.
2. Brument, N., Morenweiser, R., Blouin, V., Toublanc, E., Raimbaud, I. et al. (2002) *Mol Ther* **6**:678–86.
3. Clark, K., Liu, X., McGrath, J., and Johnson, P. (1999) *Hum. Gene Ther.*, **10**, 1031-1039.
4. Graham, F. L., Smiley, J., Russell, W. C. and Nairn, R. (1977) *J Gen Virol* **36**:59-74.
5. Grimm, D. and Kleinschmidt, J. A. (1999) *Hum Gene Ther* **10**:2445-50.
6. Matsushita, T., Elliger, S., Elliger, C., Podsakoff, G., Villarreal, L. et al. (1998) *Gene Ther* **5**:938-45.

7. McCarty, D. M., Monahan, P. E. and Sumulski, R. J. (2001) *Gene Therapy* **8**:1248-1254.
8. Rabinowitz, J, and Samulski, R. J. (1998) *Curr. Opin. Biotechnol.*, **9**, 470-475.
9. Russell, D. W., Alexander, I. E. and Miller, A. D. (1995) *Proc Natl Acad Sci U S A* **92**:5719-23.
10. Summer ford, C., and Samulski, R. J. (1999) *Nat. Med.*, **5**, 587-588.

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1. Kim, K.L. et al. (2026). FGF12 induces aberrant mechanosignaling in aortic smooth muscle cells during thoracic aortic aneurysm formation in Marfan syndrome mice. *Exp Mol Med*. doi: 10.1038/s12276-025-01621-y.
2. Lee, H.Y. et al. (2025). Astrocytic monoamine oxidase B (MAOB)-gamma-aminobutyric acid (GABA) axis as a molecular brake on repair following spinal cord injury. *Signal Transduct Target Ther*. **10**(1):295. doi: 10.1038/s41392-025-02398-2.
3. Lin, Y.L. et al. (2025). RNF128 regulates the adaptive metabolic response to fasting by modulating PPAR α function. *Cell Death Differ*. doi: 10.1038/s41418-025-01579-4.
4. Wang, B. et al. (2025). Lysophosphatidic acid receptor 5 in insular cortex as a potential analgesic target in neuropathic pain. *Neurotherapeutics*. doi: 10.1016/j.neurot.2025.e00609.
5. Tsai, Y.W. et al. (2025). N-Cadherin promotes cardiac regeneration by potentiating pro-mitotic β -Catenin signaling in cardiomyocytes. *Nat Commun*. **16**(1):896. doi: 10.1038/s41467-025-56216-y.
6. Kim, Y.J. et al. (2024). Synthetic ShK-like Peptide from the Jellyfish *Nemopilema nomurai* Has Human Voltage-Gated Potassium-Channel-Blocking Activity. *Mar Drugs*. **22**(5):217. doi: 10.3390/md22050217.
7. Kim, H.J. et al. (2024). ROR α -GABP-TFAM axis alleviates myosteatosis with fatty atrophy through reinforcement of mitochondrial capacity. *J Cachexia Sarcopenia Muscle*. **15**(2):615-630. doi: 10.1002/jcsm.13432.
8. Tsuyama, T. et al. (2023). Hypoxia causes pancreatic β -cell dysfunction and impairs insulin secretion by activating the transcriptional repressor BHLHE40. *EMBO Rep*. doi: 10.15252/embr.202256227.

9. Liu, P.Y. et al. (2023). RNF128 regulates neutrophil infiltration and myeloperoxidase functions to prevent acute lung injury. *Cell Death Dis.* **14**(6):369. doi: 10.1038/s41419-023-05890-1.
10. Morioka, S. et al. (2022). Chimeric efferocytic receptors improve apoptotic cell clearance and alleviate inflammation. *Cell.* **185**(26):4887-4903.e17. doi: 10.1016/j.cell.2022.11.029.
11. Yan, H. et al. (2022). MDMX elevation by a novel Mdmx-p53 interaction inhibitor mitigates neuronal damage after ischemic stroke. *Sci Rep.* **12**(1):21110. doi: 10.1038/s41598-022-25427-4.
12. Seo, J. et al. (2022). Myristoylation-dependent palmitoylation of cyclin Y modulates long-term potentiation and spatial learning. *Prog Neurobiol.* doi: 10.1016/j.pneurobio.2022.102349.
13. Tricaud, N. et al. (2022). Traumatic and Diabetic Schwann Cell Demyelination Is Triggered by a Transient Mitochondrial Calcium Release through Voltage Dependent Anion Channel 1. *Biomedicines.* **10**(6):1447. doi: 10.3390/biomedicines10061447.
14. Boyer, J.G. et al. (2022). Depletion of skeletal muscle satellite cells attenuates pathology in muscular dystrophy. *Nat Commun.* **13**(1):2940. doi: 10.1038/s41467-022-30619-7.
15. Kashihara, T. et al. (2022). YAP mediates compensatory cardiac hypertrophy through aerobic glycolysis in response to pressure overload. *J Clin Invest.* doi: 10.1172/JCI150595.
16. Huang, C. et al. (2021). TBN improves motor function and prolongs survival in a TDP-43M337V mouse model of ALS. *Hum Mol Genet.* doi: 10.1093/hmg/ddab101.
17. Zubkova, E.S. et al. (2021). Transduction of rat and human adipose-tissue derived mesenchymal stromal cells by adeno-associated viral vector serotype DJ. *Biol Open.* doi: 10.1242/bio.058461.
18. Vanhoutte, D. et al. (2021). Thbs1 induces lethal cardiac atrophy through PERK-ATF4 regulated autophagy. *Nat Commun.* **12**(1):3928. doi: 10.1038/s41467-021-24215-4.
19. Asahina, M. et al. (2021). Reversibility of motor dysfunction in the rat model of NGLY1 deficiency. *Mol Brain.* **14**(1):91. doi: 10.1186/s13041-021-00806-6.
20. Liu, P.Y. et al. (2021). E3 ubiquitin ligase Grail promotes hepatic steatosis through Sirt1 inhibition. *Cell Death Dis.* **12**(4):323. doi: 10.1038/s41419-021-03608-9.
21. Bai, M. et al. (2020). Critical regulation of a NDIME/MEF2C axis in embryonic stem cell neural differentiation and autism. *EMBO Rep.* doi: 10.15252/embr.202050283.
22. Silva-Pilipich, N. et al. (2020). Long-Term Systemic Expression of a Novel PD-1 Blocking Nanobody from an AAV Vector Provides Antitumor Activity without Toxicity. *Biomedicines.* **8**(12):E562. doi: 10.3390/biomedicines8120562.
23. Zhu, J. et al. (2020). Preparation of a Bacteriophage T4-based Prokaryotic-eukaryotic Hybrid Viral Vector for Delivery of Large Cargos of Genes and Proteins into Human Cells. *Bio-protocol.* **10**(07): e3573. doi: 10.21769/BioProtoc.3573.
24. Deng, X. et al. (2020). Establishment of a Recombinant AAV2/HBoV1 Vector Production System in Insect Cells. *Genes (Basel).* **11**(4). pii: E439. doi: 10.3390/genes11040439.
25. Jang, M. et al. (2018). Adeno-Associated Viral Vector Serotype DJ-Mediated Overexpression of N171-82Q-Mutant Huntingtin in the Striatum of Juvenile Mice Is a New Model for Huntington's Disease. *Front Cell Neurosci.* **12**:157. doi: 10.3389/fncel.2018.00157.
26. Hoshino, Y. et al. (2019). The adeno-associated virus rh10 vector is an effective gene transfer system for chronic spinal cord injury. *Sci Rep.* **9**(1):9844. doi: 10.1038/s41598-019-46069-z.
27. van Hameren, G. et al. (2019). In vivo real-time dynamics of ATP and ROS production in axonal mitochondria show decoupling in mouse models of peripheral neuropathies. *Acta Neuropathol Commun.* **7**(1):13. doi: 10.1186/s40478-019-0740-4.
28. Andrzejewski, S. et al. (2019). Adeno-associated virus neutralising antibodies in type 1 diabetes mellitus. *Gene Ther.* doi: 10.1038/s41434-019-0076-5.

29. Jang, M. et al. (2019). Gintonin, a ginseng-derived ingredient, as a novel therapeutic strategy for Huntington's disease: Activation of the Nrf2 pathway through lysophosphatidic acid receptors. *Brain Behav Immun.* pii: S0889-1591(18)30392-1. doi: 10.1016/j.bbi.2019.03.001.
30. Nakamura, M. et al. (2019). Glycogen Synthase Kinase-3 α Promotes Fatty Acid Uptake and Lipotoxic Cardiomyopathy. *Cell Metab.* pii: S1550-4131(19)30005-1. doi: 10.1016/j.cmet.2019.01.005.

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